

# VU Research Portal

## Advanced Imaging in Glioma Treatment

Verburg, N.

2020

### **document version**

Publisher's PDF, also known as Version of record

[Link to publication in VU Research Portal](#)

### **citation for published version (APA)**

Verburg, N. (2020). *Advanced Imaging in Glioma Treatment: Moving the Frontier*. [PhD-Thesis - Research and graduation internal, Vrije Universiteit Amsterdam].

### **General rights**

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

### **Take down policy**

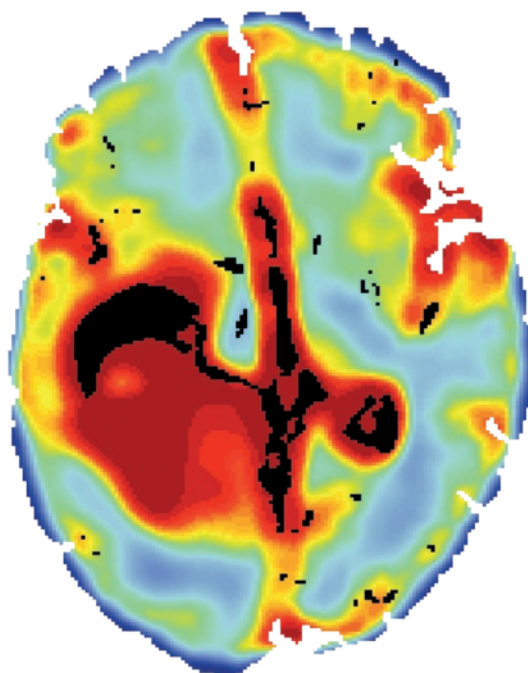
If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

### **E-mail address:**

[vuresearchportal.ub@vu.nl](mailto:vuresearchportal.ub@vu.nl)



## Appendices



# I. References

1. Ho VK, Reijneveld JC, Enting RH, et al. Changing incidence and improved survival of gliomas. *European journal of cancer* 2014;50:2309-18.
2. Weller M, van den Bent M, Tonn JC, et al. European Association for Neuro-Oncology (EANO) guideline on the diagnosis and treatment of adult astrocytic and oligodendroglial gliomas. *The Lancet Oncology* 2017;18:e315-e29.
3. Kelly PJ, Daumas-Duport C, Kispert DB, Kall BA, Scheithauer BW, Illig JJ. Imaging-based stereotaxic serial biopsies in untreated intracranial glial neoplasms. *Journal of neurosurgery* 1987;66:865-74.
4. Pallud J, Varlet P, Devaux B, et al. Diffuse low-grade oligodendrogliomas extend beyond MRI-defined abnormalities. *Neurology* 2010;74:1724-31.
5. Döbelbower MC, Burnett III OL, Nordal RA, et al. Patterns of failure for glioblastoma multiforme following concurrent radiation and temozolomide. *Journal of medical imaging and radiation oncology* 2011;55:77-81.
6. McDonald MW, Shu HK, Curran WJ, Jr., Crocker IR. Pattern of failure after limited margin radiotherapy and temozolomide for glioblastoma. *International journal of radiation oncology, biology, physics* 2011;79:130-6.
7. Li YM, Suki D, Hess K, Sawaya R. The influence of maximum safe resection of glioblastoma on survival in 1229 patients: Can we do better than gross-total resection? *Journal of neurosurgery* 2016;124:977-88.
8. Smith JS, Chang EF, Lamborn KR, et al. Role of extent of resection in the long-term outcome of low-grade hemispheric gliomas. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2008;26:1338-45.
9. Louis DN, Perry A, Reifenberger G, et al. The 2016 World Health Organization Classification of Tumors of the Central Nervous System: a summary. *Acta neuropathologica* 2016;131:803-20.
10. Ostrom QT, Gittleman H, Truitt G, Boscia A, Kruchko C, Barnholtz-Sloan JS. CBTRUS Statistical Report: Primary Brain and Other Central Nervous System Tumors Diagnosed in the United States in 2011-2015. *Neuro-oncology* 2018;20:iv1-iv86.
11. Miranda-Filho A, Pineros M, Soerjomataram I, Deltour I, Bray F. Cancers of the brain and CNS: global patterns and trends in incidence. *Neuro-oncology* 2017;19:270-80.
12. Silversmit G, Vaes E, van Eycken L. Estimation of population-based cancer-specific potential years of life lost in Belgium. *European journal of cancer prevention : the official journal of the European Cancer Prevention Organisation* 2017;26:Joining forces for better cancer registration in Europe:S157-S63.
13. Barres BA. The mystery and magic of glia: a perspective on their roles in health and disease. *Neuron* 2008;60:430-40.
14. Sanai N, Alvarez-Buylla A, Berger MS. Neural stem cells and the origin of gliomas. *The New England journal of medicine* 2005;353:811-22.



15. Eckel-Passow JE, Lachance DH, Molinaro AM, et al. Glioma Groups Based on 1p/19q, IDH, and TERT Promoter Mutations in Tumors. *The New England journal of medicine* 2015;372:2499-508.
16. Turcan S, Rohle D, Goenka A, et al. IDH1 mutation is sufficient to establish the glioma hypermethylator phenotype. *Nature* 2012;483:479-83.
17. Dor Y, Cedar H. Principles of DNA methylation and their implications for biology and medicine. *Lancet* 2018;392:777-86.
18. Brat DJ, Aldape K, Colman H, et al. cIMPACT-NOW update 3: recommended diagnostic criteria for "Diffuse astrocytic glioma, IDH-wildtype, with molecular features of glioblastoma, WHO grade IV". *Acta neuropathologica* 2018;136:805-10.
19. Stichel D, Ebrahimi A, Reuss D, et al. Distribution of EGFR amplification, combined chromosome 7 gain and chromosome 10 loss, and TERT promoter mutation in brain tumors and their potential for the reclassification of IDHwt astrocytoma to glioblastoma. *Acta neuropathologica* 2018;136:793-803.
20. Barthel FP, Wesseling P, Verhaak RGW. Reconstructing the molecular life history of gliomas. *Acta neuropathologica* 2018;135:649-70.
21. Wesseling P, Capper D. WHO 2016 Classification of gliomas. *Neuropathology and applied neurobiology* 2018;44:139-50.
22. Bai H, Harmanci AS, Erson-Omay EZ, et al. Integrated genomic characterization of IDH1-mutant glioma malignant progression. *Nature genetics* 2016;48:59-66.
23. Cohen A, Sato M, Aldape K, et al. DNA copy number analysis of Grade II-III and Grade IV gliomas reveals differences in molecular ontogeny including chromothripsis associated with IDH mutation status. *Acta neuropathologica communications* 2015;3:34.
24. Watkins S, Robel S, Kimbrough IF, Robert SM, Ellis-Davies G, Sontheimer H. Disruption of astrocyte-vascular coupling and the blood-brain barrier by invading glioma cells. *Nature communications* 2014;5:4196.
25. Jain RK, di Tomaso E, Duda DG, Loeffler JS, Sorensen AG, Batchelor TT. Angiogenesis in brain tumours. *Nature reviews Neuroscience* 2007;8:610-22.
26. Giese A, Bjerkvig R, Berens ME, Westphal M. Cost of migration: invasion of malignant gliomas and implications for treatment. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2003;21:1624-36.
27. Willard N, Kleinschmidt-DeMasters BK. Massive dissemination of adult glioblastomas. *Clinical neuropathology* 2015;34:330-42.
28. Louis DN, Ohgaki H, Wiestler OD, et al. The 2007 WHO classification of tumours of the central nervous system. *Acta neuropathologica* 2007;114:97-109.
29. Price SJ, Gillard JH. Imaging biomarkers of brain tumour margin and tumour invasion. *The British journal of radiology* 2011;84 Spec No 2:S159-67.
30. Sahm F, Capper D, Jeibmann A, et al. Addressing diffuse glioma as a systemic brain disease with single-cell analysis. *Archives of neurology* 2012;69:523-6.
31. Stensjoen AL, Solheim O, Kvistad KA, Haberg AK, Salvesen O, Berntsen EM. Growth dynamics of untreated glioblastomas in vivo. *Neuro-oncology* 2015;17:1402-11.

32. Gui C, Kosteniuk SE, Lau JC, Megyesi JF. Tumor growth dynamics in serially-imaged low-grade glioma patients. *Journal of neuro-oncology* 2018;139:167-75.
33. Rossetti AO, Stupp R. Epilepsy in brain tumor patients. *Current opinion in neurology* 2010;23:603-9.
34. Karnofsky DA, Burchenal JH. The clinical evaluation of chemotherapeutic agents in cancer. In: Macloed CM, ed. *Evaluation of chemo-therapeutic Agents*. New York: Columbia University Press; 1949:191-205.
35. Lamborn KR, Chang SM, Prados MD. Prognostic factors for survival of patients with glioblastoma: recursive partitioning analysis. *Neuro-oncology* 2004;6:227-35.
36. Potts MB, Smith JS, Molinaro AM, Berger MS. Natural history and surgical management of incidentally discovered low-grade gliomas. *Journal of neurosurgery* 2012;116:365-72.
37. Pallud J, Fontaine D, Duffau H, et al. Natural history of incidental World Health Organization grade II gliomas. *Annals of neurology* 2010;68:727-33.
38. Ellingson BM, Bendszus M, Boxerman J, et al. Consensus recommendations for a standardized Brain Tumor Imaging Protocol in clinical trials. *Neuro-oncology* 2015;17:1188-98.
39. Scott JN, Brasher PM, Sevic RJ, Rewcastle NB, Forsyth PA. How often are nonenhancing supratentorial gliomas malignant? A population study. *Neurology* 2002;59:947-9.
40. Burger PC, Scheithauer BW. *Tumours of the Central Nervous System (Afip Atlas of Tumour Pathology)* (4<sup>th</sup> ed.). Washington: American Registry of Pathology; 2007:34.
41. Smits M. Imaging of oligodendroglioma. *The British journal of radiology* 2016;89:20150857.
42. Diffuse astrocytoma. at [https://radiopaedia.org/articles/diffuse-astrocytoma-1?lang=us.](https://radiopaedia.org/articles/diffuse-astrocytoma-1?lang=us)
43. Brown TJ, Brennan MC, Li M, et al. Association of the Extent of Resection With Survival in Glioblastoma: A Systematic Review and Meta-analysis. *JAMA oncology* 2016;2:1460-9.
44. Niyazi M, Brada M, Chalmers AJ, et al. ESTRO-ACROP guideline “target delineation of glioblastomas”. *Radiotherapy and oncology : journal of the European Society for Therapeutic Radiology and Oncology* 2016;118:35-42.
45. Whitfield GA, Kennedy SR, Djoukharadar IK, Jackson A. Imaging and target volume delineation in glioma. *Clinical oncology* 2014;26:364-76.
46. van den Bent MJ, Afra D, de Witte O, et al. Long-term efficacy of early versus delayed radiotherapy for low-grade astrocytoma and oligodendroglioma in adults: the EORTC 22845 randomised trial. *Lancet* 2005;366:985-90.
47. Laperriere N, Zuraw L, Cairncross G, Cancer Care Ontario Practice Guidelines Initiative Neuro-Oncology Disease Site G. Radiotherapy for newly diagnosed malignant glioma in adults: a systematic review. *Radiotherapy and oncology : journal of the European Society for Therapeutic Radiology and Oncology* 2002;64:259-73.
48. Buckner JC, Shaw EG, Pugh SL, et al. Radiation plus Procarbazine, CCNU, and Vincristine in Low-Grade Glioma. *The New England journal of medicine* 2016;374:1344-55.
49. Stupp R, Mason WP, van den Bent MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *The New England journal of medicine* 2005;352:987-96.

50. Stummer W, Pichlmeier U, Meinel T, et al. Fluorescence-guided surgery with 5-aminolevulinic acid for resection of malignant glioma: a randomised controlled multicentre phase III trial. *The Lancet Oncology* 2006;7:392-401.
51. Ewelt C, Floeth FW, Felsberg J, et al. Finding the anaplastic focus in diffuse gliomas: the value of Gd-DTPA enhanced MRI, FET-PET, and intraoperative, ALA-derived tissue fluorescence. *Clinical neurology and neurosurgery* 2011;113:541-7.
52. Floeth FW, Sabel M, Ewelt C, et al. Comparison of (18)F-FET PET and 5-ALA fluorescence in cerebral gliomas. *European journal of nuclear medicine and molecular imaging* 2011;38:731-41.
53. Price SJ, Jena R, Burnet NG, et al. Improved delineation of glioma margins and regions of infiltration with the use of diffusion tensor imaging: an image-guided biopsy study. *AJNR American journal of neuroradiology* 2006;27:1969-74.
54. Pauleit D, Floeth F, Hamacher K, et al. O-(2-[18F]fluoroethyl)-L-tyrosine PET combined with MRI improves the diagnostic assessment of cerebral gliomas. *Brain : a journal of neurology* 2005;128:678-87.
55. Kuhn SA, Romeike B, Walter J, Kalff R, Reichart R. Multiplanar MRI-CT fusion neuronavigation-guided serial stereotactic biopsy of human brain tumors: proof of a strong correlation between tumor imaging and histopathology by a new technical approach. *Journal of cancer research and clinical oncology* 2009;135:1293-302.
56. Becker G, Hofmann E, Woydt M, et al. Postoperative neuroimaging of high-grade gliomas: comparison of transcranial sonography, magnetic resonance imaging, and computed tomography. *Neurosurgery* 1999;44:469-77; discussion 77-8.
57. Hermann EJ, Hattingen E, Krauss JK, et al. Stereotactic biopsy in gliomas guided by 3-tesla 1H-chemical-shift imaging of choline. *Stereotactic and functional neurosurgery* 2008;86:300-7.
58. McGirt MJ, Villavicencio AT, Bulsara KR, Friedman AH. MRI-guided stereotactic biopsy in the diagnosis of glioma: comparison of biopsy and surgical resection specimen. *Surgical neurology* 2003;59:277-81; discussion 81-2.
59. Jain R, Gutierrez J, Narang J, et al. In vivo correlation of tumor blood volume and permeability with histologic and molecular angiogenic markers in gliomas. *AJNR American journal of neuroradiology* 2011;32:388-94.
60. Panciani PP, Fontanella M, Schatlo B, et al. Fluorescence and image guided resection in high grade glioma. *Clinical neurology and neurosurgery* 2012;114:37-41.
61. Widhalm G, Minchev G, Woehrer A, et al. Strong 5-aminolevulinic acid-induced fluorescence is a novel intraoperative marker for representative tissue samples in stereotactic brain tumor biopsies. *Neurosurgical review* 2012;35:381-91; discussion 91.
62. Yamaguchi S, Kobayashi H, Hirata K, et al. Detection of histological anaplasia in gliomas with oligodendroglial components using positron emission tomography with (18)F-FDG and (11) C-methionine: report of two cases. *Journal of neuro-oncology* 2011;101:335-41.
63. Barajas RF, Jr., Phillips JJ, Parvataneni R, et al. Regional variation in histopathologic features of tumor specimens from treatment-naïve glioblastoma correlates with anatomic and physiologic MR Imaging. *Neuro-oncology* 2012;14:942-54.

64. Guo J, Yao C, Chen H, et al. The relationship between Cho/NAA and glioma metabolism: implementation for margin delineation of cerebral gliomas. *Acta neurochirurgica* 2012;154:1361-70; discussion 70.
65. Barajas RF, Jr., Hess CP, Phillips JJ, et al. Super-resolution track density imaging of glioblastoma: histopathologic correlation. *AJNR American journal of neuroradiology* 2013;34:1319-25.
66. Gill BJ, Pisapia DJ, Malone HR, et al. MRI-localized biopsies reveal subtype-specific differences in molecular and cellular composition at the margins of glioblastoma. *Proceedings of the National Academy of Sciences of the United States of America* 2014;111:12550-5.
67. Kamson DO, Juhasz C, Buth A, et al. Tryptophan PET in pretreatment delineation of newly-diagnosed gliomas: MRI and histopathologic correlates. *Journal of neuro-oncology* 2013;112:121-32.
68. Pafundi DH, Laack NN, Youland RS, et al. Biopsy validation of 18F-DOPA PET and biodistribution in gliomas for neurosurgical planning and radiotherapy target delineation: results of a prospective pilot study. *Neuro-oncology* 2013;15:1058-67.
69. Shinomiya A, Kawai N, Okada M, et al. Evaluation of 3'-deoxy-3'-[18F]-fluorothymidine (18F-FLT) kinetics correlated with thymidine kinase-1 expression and cell proliferation in newly diagnosed gliomas. *European journal of nuclear medicine and molecular imaging* 2013;40:175-85.
70. Watanabe M, Tanaka R, Takeda N. Magnetic resonance imaging and histopathology of cerebral gliomas. *Neuroradiology* 1992;34:463-9.
71. Pamir MN, Ozduman K, Yildiz E, Sav A, Dincer A. Intraoperative magnetic resonance spectroscopy for identification of residual tumor during low-grade glioma surgery: clinical article. *Journal of neurosurgery* 2013;118:1191-8.
72. Surov A, Meyer HJ, Wienke A. Correlation between apparent diffusion coefficient (ADC) and cellularity is different in several tumors: a meta-analysis. *Oncotarget* 2017;8:59492-9.
73. Pauleit D, Langen KJ, Floeth F, et al. Can the apparent diffusion coefficient be used as a noninvasive parameter to distinguish tumor tissue from peritumoral tissue in cerebral gliomas? *Journal of magnetic resonance imaging : JMRI* 2004;20:758-64.
74. Li Y, Zhang W. Quantitative evaluation of diffusion tensor imaging for clinical management of glioma. *Neurosurgical review* 2018.
75. Petrella JR, Provenzale JM. MR perfusion imaging of the brain: techniques and applications. *AJR American journal of roentgenology* 2000;175:207-19.
76. Jain R, Griffith B, Alotaibi F, et al. Glioma Angiogenesis and Perfusion Imaging: Understanding the Relationship between Tumor Blood Volume and Leakiness with Increasing Glioma Grade. *AJNR American journal of neuroradiology* 2015;36:2030-5.
77. Zhu H, Barker PB. MR spectroscopy and spectroscopic imaging of the brain. *Methods in molecular biology* 2011;711:203-26.
78. Glunde K, Bhujwala ZM, Ronen SM. Choline metabolism in malignant transformation. *Nature reviews Cancer* 2011;11:835-48.
79. Zhang J, Zhuang DX, Yao CJ, et al. Metabolic approach for tumor delineation in glioma surgery: 3D MR spectroscopy image-guided resection. *Journal of neurosurgery* 2016;124:1585-93.

80. New PF, Scott WR, Schnur JA, Davis KR, Taveras JM, Hochberg FH. Computed tomography with the EMI scanner in the diagnosis of primary and metastatic intracranial neoplasms. *Radiology* 1975;114:75-87.
81. Sastry R, Bi WL, Pieper S, et al. Applications of Ultrasound in the Resection of Brain Tumors. *Journal of neuroimaging : official journal of the American Society of Neuroimaging* 2017;27:5-15.
82. Brindle KM, Izquierdo-Garcia JL, Lewis DY, Mair RJ, Wright AJ. Brain Tumor Imaging. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2017;35:2432-8.
83. Albert NL, Weller M, Suchorska B, et al. Response Assessment in Neuro-Oncology working group and European Association for Neuro-Oncology recommendations for the clinical use of PET imaging in gliomas. *Neuro-oncology* 2016;18:1199-208.
84. Langen K-J, Galldiks N, Hattingen E, Shah NJ. Advances in neuro-oncology imaging. *Nature reviews Neurology* 2017.
85. Orringer DA, Golby A, Jolesz F. Neuronavigation in the surgical management of brain tumors: current and future trends. *Expert review of medical devices* 2012;9:491-500.
86. Widmann G, Schullian P, Ortler M, Bale R. Frameless stereotactic targeting devices: technical features, targeting errors and clinical results. *The international journal of medical robotics + computer assisted surgery : MRCAS* 2012;8:1-16.
87. Ricard D, Idhahbi A, Ducray F, Lahutte M, Hoang-Xuan K, Delattre JY. Primary brain tumours in adults. *Lancet* 2012;379:1984-96.
88. van den Bent MJ, Wefel JS, Schiff D, et al. Response assessment in neuro-oncology (a report of the RANO group): assessment of outcome in trials of diffuse low-grade gliomas. *The Lancet Oncology* 2011;12:583-93.
89. Wen PY, Macdonald DR, Reardon DA, et al. Updated response assessment criteria for high-grade gliomas: response assessment in neuro-oncology working group. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2010;28:1963-72.
90. De Witt Hamer PC, Robles SG, Zwinderman AH, Duffau H, Berger MS. Impact of intraoperative stimulation brain mapping on glioma surgery outcome: a meta-analysis. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2012;30:2559-65.
91. Capelle L, Fontaine D, Mandonnet E, et al. Spontaneous and therapeutic prognostic factors in adult hemispheric World Health Organization Grade II gliomas: a series of 1097 cases: clinical article. *Journal of neurosurgery* 2013;118:1157-68.
92. Einstein DB, Wessels B, Bangert B, et al. Phase II trial of radiosurgery to magnetic resonance spectroscopy-defined high-risk tumor volumes in patients with glioblastoma multiforme. *International journal of radiation oncology, biology, physics* 2012;84:668-74.
93. Sanai N, Polley MY, McDermott MW, Parsa AT, Berger MS. An extent of resection threshold for newly diagnosed glioblastomas. *Journal of neurosurgery* 2011;115:3-8.
94. Pichlmeier U, Bink A, Schackert G, Stummer W. Resection and survival in glioblastoma multiforme: an RTOG recursive partitioning analysis of ALA study patients. *Neuro-oncology* 2008;10:1025-34.

95. la Fougère C, Suchorska B, Bartenstein P, Kreth F-W, Tonn J-C. Molecular imaging of gliomas with PET: opportunities and limitations. *Neuro-oncology* 2011;13:806-19.
96. Waldman AD, Jackson A, Price SJ, et al. Quantitative imaging biomarkers in neuro-oncology. *Nat Rev Clin Oncol* 2009;6:445-54.
97. Landis JR, Koch GG. The measurement of observer agreement for categorical data. *Biometrics* 1977;33:159-74.
98. Bossuyt PM, Reitsma JB, Bruns DE, et al. STARD 2015: an updated list of essential items for reporting diagnostic accuracy studies. *Bmj* 2015;351:h5527.
99. Rutter CM, Gatsonis CA. A hierarchical regression approach to meta-analysis of diagnostic test accuracy evaluations. *Stat Med* 2001;20:2865-84.
100. Glas AS, Lijmer JG, Prins MH, Bossel GJ, Bossuyt PMM. The diagnostic odds ratio: a single indicator of test performance. *Journal of Clinical Epidemiology* 2003;56:1129-35.
101. Macaskill P. Empirical Bayes estimates generated in a hierarchical summary ROC analysis agreed closely with those of a full Bayesian analysis. *J Clin Epidemiol* 2004;57:925-32.
102. Harbord RM, Whiting P, Sterne JAC, et al. An empirical comparison of methods for meta-analysis of diagnostic accuracy showed hierarchical models are necessary. *Journal of Clinical Epidemiology* 2008;61:1095-103.
103. Riley RD, Dodd SR, Craig JV, Thompson JR, Williamson PR. Meta-analysis of diagnostic test studies using individual patient data and aggregate data. *Stat Med* 2008;27:6111-36.
104. Sutton AJ, Kendrick D, Coupland CA. Meta-analysis of individual- and aggregate-level data. *Stat Med* 2008;27:651-69.
105. Dukic V, Gatsonis C. Meta-analysis of diagnostic test accuracy assessment studies with varying number of thresholds. *Biometrics* 2003;59:936-46.
106. Leeflang MMG, Deeks JJ, Gatsonis C, Bossuyt PMM, Group CDTAW. Systematic reviews of diagnostic test accuracy. *Annals of internal medicine* 2008;149:889-97.
107. Rutjes AWS, Reitsma JB, Di Nisio M, Smidt N, van Rijn JC, Bossuyt PMM. Evidence of bias and variation in diagnostic accuracy studies. *CMAJ : Canadian Medical Association journal = journal de l'Association medicale canadienne* 2006;174:469-76.
108. Chou R, Cuevas C, Fu R, et al. Imaging techniques for the diagnosis of hepatocellular carcinoma: a systematic review and meta-analysis. *Annals of internal medicine* 2015;162:697-711.
109. Silvestri GA, Gonzalez AV, Jantz MA, et al. Methods for staging non-small cell lung cancer: Diagnosis and management of lung cancer, 3rd ed: American College of Chest Physicians evidence-based clinical practice guidelines. *Chest* 2013;143:e211S-50S.
110. Zhang Y, Qin Q, Li B, Wang J, Zhang K. Magnetic resonance imaging for N staging in non-small cell lung cancer: A systematic review and meta-analysis. *Thorac Cancer* 2015;6:123-32.
111. Peters NH, Borel Rinkes IH, Zuithoff NP, Mali WP, Moons KG, Peeters PH. Meta-analysis of MR imaging in the diagnosis of breast lesions. *Radiology* 2008;246:116-24.
112. Lemée J-M, Clavreul A, Aubry M, et al. Characterizing the peritumoral brain zone in glioblastoma: a multidisciplinary analysis. *Journal of neuro-oncology* 2015;122:53-61.

113. Kros JM, Gorlia T, Kouwenhoven MC, et al. Panel review of anaplastic oligodendroglioma from European Organization For Research and Treatment of Cancer Trial 26951: assessment of consensus in diagnosis, influence of 1p/19q loss, and correlations with outcome. *Journal of neuropathology and experimental neurology* 2007;66:545-51.
114. Scott CB, Nelson JS, Farnan NC, et al. Central pathology review in clinical trials for patients with malignant glioma. A report of Radiation Therapy Oncology Group 83-02. *Cancer* 1995;76:307-13.
115. van den Bent MJ. Interobserver variation of the histopathological diagnosis in clinical trials on glioma: a clinician's perspective. *Acta neuropathologica* 2010;120:297-304.
116. Bjartmarz H, Rehncrona S. Comparison of accuracy and precision between frame-based and frameless stereotactic navigation for deep brain stimulation electrode implantation. *Stereotactic and functional neurosurgery* 2007;85:235-42.
117. Zacest A, Berk C, Burchiel KJ. Precision and accuracy of stereotactic targeting in patients undergoing repeat stereotactic surgery. *Stereotactic and functional neurosurgery* 2009;87:168-73.
118. Sottoriva A, Spiteri I, Piccirillo SGM, et al. Intratumor heterogeneity in human glioblastoma reflects cancer evolutionary dynamics. *Proceedings of the National Academy of Sciences of the United States of America* 2013;110:4009-14.
119. Cabrera AR, Kirkpatrick JP, Fiveash JB, et al. Radiation therapy for glioblastoma: Executive summary of an American Society for Radiation Oncology Evidence-Based Clinical Practice Guideline. *Practical radiation oncology* 2016;6:217-25.
120. Zhao F, Li M, Kong L, Zhang G, Yu J. Delineation of radiation therapy target volumes for patients with postoperative glioblastoma: a review. *OncoTargets and therapy* 2016;9:3197-204.
121. Albert NL, Weller M, Suchorska B, et al. Response Assessment in Neuro-Oncology working group and European Association for Neuro-Oncology recommendations for the clinical use of PET imaging in gliomas. *Neuro-oncology* 2016;18:1199-208.
122. Brandão LA, Castillo M. Adult Brain Tumors: Clinical Applications of Magnetic Resonance Spectroscopy. *Magnetic resonance imaging clinics of North America* 2016;24:781-809.
123. Fink JR, Muzi M, Peck M, Krohn KA. Multimodality Brain Tumor Imaging: MR Imaging, PET, and PET/MR Imaging. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine* 2015;56:1554-61.
124. Kalpathy-Cramer J, Gerstner ER, Emblem KE, Andronesi OC, Rosen B. Advanced magnetic resonance imaging of the physical processes in human glioblastoma. *Cancer research* 2014;74:4622-37.
125. Lijmer JG, Bossuyt PMM, Heisterkamp SH. Exploring sources of heterogeneity in systematic reviews of diagnostic tests. *Statistics in medicine* 2002;21:1525-37.
126. Verburg N, Hoefnagels FWA, Barkhof F, et al. Diagnostic Accuracy of Neuroimaging to Delineate Diffuse Gliomas within the Brain: A Meta-Analysis. *AJNR American journal of neuroradiology* 2017;38:1884-91.

127. Keles GE, Chang EF, Lamborn KR, et al. Volumetric extent of resection and residual contrast enhancement on initial surgery as predictors of outcome in adult patients with hemispheric anaplastic astrocytoma. *Journal of neurosurgery* 2006;105:34-40.
128. Pessina F, Navarria P, Cozzi L, et al. Maximize surgical resection beyond contrast-enhancing boundaries in newly diagnosed glioblastoma multiforme: is it useful and safe? A single institution retrospective experience. *Journal of neuro-oncology* 2017;135:129-39.
129. Pirotte B, Goldman S, Dewitte O, et al. Integrated positron emission tomography and magnetic resonance imaging-guided resection of brain tumors: a report of 103 consecutive procedures. *Journal of neurosurgery* 2006;104:238-53.
130. Pirotte BJ, Levivier M, Goldman S, et al. Positron emission tomography-guided volumetric resection of supratentorial high-grade gliomas: a survival analysis in 66 consecutive patients. *Neurosurgery* 2009;64:471-81; discussion 81.
131. Ening G, Osterheld F, Capper D, Schmieder K, Brenke C. Risk factors for glioblastoma therapy associated complications. *Clinical neurology and neurosurgery* 2015;134:55-9.
132. Aghi MK, Nahed BV, Sloan AE, Ryken TC, Kalkanis SN, Olson JJ. The role of surgery in the management of patients with diffuse low grade glioma: A systematic review and evidence-based clinical practice guideline. *Journal of neuro-oncology* 2015;125:503-30.
133. Ius T, Isola M, Budai R, et al. Low-grade glioma surgery in eloquent areas: volumetric analysis of extent of resection and its impact on overall survival. A single-institution experience in 190 patients: clinical article. *Journal of neurosurgery* 2012;117:1039-52.
134. Henker C, Hiepel MC, Kriesen T, et al. Volumetric assessment of glioblastoma and its predictive value for survival. *Acta neurochirurgica* 2019;161:1723-32.
135. Kelly PJ, Daumas-Duport C, Scheithauer BW, Kall BA, Kispert DB. Stereotactic histologic correlations of computed tomography- and magnetic resonance imaging-defined abnormalities in patients with glial neoplasms. *Mayo Clinic proceedings* 1987;62:450-9.
136. Li Y, Osorio JA, Ozturk-Isik E, et al. Considerations in applying 3D PRESS H-1 brain MRSI with an eight-channel phased-array coil at 3 T. *Magnetic resonance imaging* 2006;24:1295-302.
137. Keppler JS, Conti PS. A cost analysis of positron emission tomography. *AJR American journal of roentgenology* 2001;177:31-40.
138. Heinzel A, Stock S, Langen KJ, Muller D. Cost-effectiveness analysis of amino acid PET-guided surgery for supratentorial high-grade gliomas. *J Nucl Med* 2012;53:552-8.
139. Langen K-J, Stoffels G, Filss C, et al. Imaging of amino acid transport in brain tumours: Positron emission tomography with O-(2-[18F]fluoroethyl)-L-tyrosine (FET). *Methods* 2017;130:124-34.
140. Kubben PL, Postma AA, Kessels AG, van Overbeeke JJ, van Santbrink H. Intraobserver and interobserver agreement in volumetric assessment of glioblastoma multiforme resection. *Neurosurgery* 2010;67:1329-34.
141. Visser M, Muller DMJ, van Duijn RJM, et al. Inter-rater agreement in glioma segmentations on longitudinal MRI. *NeuroImage Clinical* 2019;22:101727.



142. Ben Abdallah M, Blonski M, Wantz-Mezieres S, Gaudeau Y, Taillandier L, Moureaux JM. Statistical evaluation of manual segmentation of a diffuse low-grade glioma MRI dataset. Conference proceedings: Annual International Conference of the IEEE Engineering in Medicine and Biology Society IEEE Engineering in Medicine and Biology Society Annual Conference 2016;2016:4403-6.
143. Hutton BF, Braun M. Software for image registration: algorithms, accuracy, efficacy. *Seminars in nuclear medicine* 2003;33:180-92.
144. Steinmeier R, Rachinger J, Kaus M, Ganslandt O, Huk W, Fahlbusch R. Factors influencing the application accuracy of neuronavigation systems. *Stereotactic and functional neurosurgery* 2000;75:188-202.
145. Nimsky C, Ganslandt O, Hastreiter P, et al. Preoperative and intraoperative diffusion tensor imaging-based fiber tracking in glioma surgery. *Neurosurgery* 2005;56:130-7; discussion 8.
146. Romano A, D'Andrea G, Calabria LF, et al. Pre- and intraoperative tractographic evaluation of corticospinal tract shift. *Neurosurgery* 2011;69:696-704; discussion -5.
147. Tanaka S, Puffer RC, Hoover JM, et al. Increased frameless stereotactic accuracy with high-field intraoperative magnetic resonance imaging. *Neurosurgery* 2012;71:ons321-7; discussion ons7-8.
148. Martin C, Alexander E, 3rd, Wong T, Schwartz R, Jolesz F, Black PM. Surgical treatment of low-grade gliomas in the intraoperative magnetic resonance imager. *Neurosurgical focus* 1998;4:e8.
149. Senft C, Bink A, Franz K, Vatter H, Gasser T, Seifert V. Intraoperative MRI guidance and extent of resection in glioma surgery: a randomised, controlled trial. *The Lancet Oncology* 2011;12:997-1003.
150. Mohammadi AM, Sullivan TB, Barnett GH, et al. Use of high-field intraoperative magnetic resonance imaging to enhance the extent of resection of enhancing and nonenhancing gliomas. *Neurosurgery* 2014;74:339-48; discussion 49; quiz 49-50.
151. Nimsky C, Ganslandt O, Von Keller B, Romstock J, Fahlbusch R. Intraoperative high-field-strength MR imaging: implementation and experience in 200 patients. *Radiology* 2004;233:67-78.
152. Pamir MN, Ozduman K, Dincer A, Yildiz E, Peker S, Ozek MM. First intraoperative, shared-resource, ultrahigh-field 3-Tesla magnetic resonance imaging system and its application in low-grade glioma resection. *Journal of neurosurgery* 2010;112:57-69.
153. Kubben PL, Scholtes F, Schijns OE, et al. Intraoperative magnetic resonance imaging versus standard neuronavigation for the neurosurgical treatment of glioblastoma: A randomized controlled trial. *Surgical neurology international* 2014;5:70.
154. Makary M, Chiocca EA, Erminy N, et al. Clinical and economic outcomes of low-field intraoperative MRI-guided tumor resection neurosurgery. *Journal of magnetic resonance imaging : JMRI* 2011;34:1022-30.
155. Kubben P. ULTRA LOW-FIELD STRENGTH INTRAOPERATIVE MRI FOR GLIOBLASTOMA SURGERY: Universiteit Maastricht; 2014.

156. Wang J, Liu X, Ba YM, et al. Effect of sonographically guided cerebral glioma surgery on survival time. *Journal of ultrasound in medicine : official journal of the American Institute of Ultrasound in Medicine* 2012;31:757-62.
157. Prada F, Vitale V, Del Bene M, et al. Contrast-enhanced MR Imaging versus Contrast-enhanced US: A Comparison in Glioblastoma Surgery by Using Intraoperative Fusion Imaging. *Radiology* 2017;285:242-9.
158. Coburger J, Konig RW, Scheuerle A, et al. Navigated high frequency ultrasound: description of technique and clinical comparison with conventional intracranial ultrasound. *World neurosurgery* 2014;82:366-75.
159. Coburger J, Scheuerle A, Kapapa T, et al. Sensitivity and specificity of linear array intraoperative ultrasound in glioblastoma surgery: a comparative study with high field intraoperative MRI and conventional sector array ultrasound. *Neurosurgical review* 2015;38:499-509; discussion
160. Prada F, Del Bene M, Mattei L, et al. Fusion imaging for intra-operative ultrasound-based navigation in neurosurgery. *Journal of ultrasound* 2014;17:243-51.
161. Le Bihan D, Jezard P, Haxby J, Sadato N, Rueckert L, Mattay V. Functional magnetic resonance imaging of the brain. *Ann Intern Med* 1995;122:296-303.
162. Giussani C, Roux FE, Ojemann J, Sganzerla EP, Pirillo D, Papagno C. Is preoperative functional magnetic resonance imaging reliable for language areas mapping in brain tumor surgery? Review of language functional magnetic resonance imaging and direct cortical stimulation correlation studies. *Neurosurgery* 2010;66:113-20.
163. Jack CR, Jr., Thompson RM, Butts RK, et al. Sensory motor cortex: correlation of presurgical mapping with functional MR imaging and invasive cortical mapping. *Radiology* 1994;190:85-92.
164. Krings T, Schreckenberger M, Rohde V, et al. Functional MRI and 18F FDG-positron emission tomography for presurgical planning: comparison with electrical cortical stimulation. *Acta neurochirurgica* 2002;144:889-99; discussion 99.
165. Roux FE, Boulanouar K, Ranjeva JP, et al. Usefulness of motor functional MRI correlated to cortical mapping in Rolandic low-grade astrocytomas. *Acta neurochirurgica* 1999;141:71-9.
166. Bartos R, Jech R, Vymazal J, et al. Validity of primary motor area localization with fMRI versus electric cortical stimulation: a comparative study. *Acta neurochirurgica* 2009;151:1071-80.
167. Bizzi A, Blasi V, Falini A, et al. Presurgical functional MR imaging of language and motor functions: validation with intraoperative electrocortical mapping. *Radiology* 2008;248:579-89.
168. Poldrack RA, Baker CI, Durnez J, et al. Scanning the horizon: towards transparent and reproducible neuroimaging research. *Nature reviews Neuroscience* 2017;18:115-26.
169. Mandonnet E, Duffau H. Chapter 5. In: Moliterno Gunel J, Piepmeyer JM, Baehring JM, eds. *Malignant Brain Tumors: State-of-the-Art Treatment* 2017.
170. Nucifora PG, Verma R, Lee SK, Melhem ER. Diffusion-tensor MR imaging and tractography: exploring brain microstructure and connectivity. *Radiology* 2007;245:367-84.

171. Berman JI, Berger MS, Chung SW, Nagarajan SS, Henry RG. Accuracy of diffusion tensor magnetic resonance imaging tractography assessed using intraoperative subcortical stimulation mapping and magnetic source imaging. *Journal of neurosurgery* 2007;107:488-94.
172. Zolal A, Hejcl A, Vachata P, et al. The use of diffusion tensor images of the corticospinal tract in intrinsic brain tumor surgery: a comparison with direct subcortical stimulation. *Neurosurgery* 2012;71:331-40; discussion 40.
173. Wu JS, Zhou LF, Tang WJ, et al. Clinical evaluation and follow-up outcome of diffusion tensor imaging-based functional neuronavigation: a prospective, controlled study in patients with gliomas involving pyramidal tracts. *Neurosurgery* 2007;61:935-48; discussion 48-9.
174. Berman JI, Chung S, Mukherjee P, Hess CP, Han ET, Henry RG. Probabilistic streamline q-ball tractography using the residual bootstrap. *NeuroImage* 2008;39:215-22.
175. Tuch DS. Q-ball imaging. *Magnetic resonance in medicine* 2004;52:1358-72.
176. Caverzasi E, Hervey-Jumper SL, Jordan KM, et al. Identifying preoperative language tracts and predicting postoperative functional recovery using HARDI q-ball fiber tractography in patients with gliomas. *Journal of neurosurgery* 2016;125:33-45.
177. Maier-Hein KH, Neher PF, Houde JC, et al. The challenge of mapping the human connectome based on diffusion tractography. *Nature communications* 2017;8:1349.
178. Wakana S, Caprihan A, Panzenboeck MM, et al. Reproducibility of quantitative tractography methods applied to cerebral white matter. *NeuroImage* 2007;36:630-44.
179. Sutherling WW, Crandall PH, Darcey TM, Becker DP, Levesque MF, Barth DS. The magnetic and electric fields agree with intracranial localizations of somatosensory cortex. *Neurology* 1988;38:1705-14.
180. Kirsch HE, Zhu Z, Honma S, Findlay A, Berger MS, Nagarajan SS. Predicting the location of mouth motor cortex in patients with brain tumors by using somatosensory evoked field measurements. *Journal of neurosurgery* 2007;107:481-7.
181. Nagarajan S, Kirsch H, Lin P, Findlay A, Honma S, Berger MS. Preoperative localization of hand motor cortex by adaptive spatial filtering of magnetoencephalography data. *Journal of neurosurgery* 2008;109:228-37.
182. Korvenoja A, Kirveskari E, Aronen HJ, et al. Sensorimotor cortex localization: comparison of magnetoencephalography, functional MR imaging, and intraoperative cortical mapping. *Radiology* 2006;241:213-22.
183. Schiffbauer H, Berger MS, Ferrari P, Freudenstein D, Rowley HA, Roberts TP. Preoperative magnetic source imaging for brain tumor surgery: a quantitative comparison with intraoperative sensory and motor mapping. *Journal of neurosurgery* 2002;97:1333-42.
184. Ganslandt O, Buchfelder M, Hastreiter P, Grummich P, Fahlbusch R, Nimsky C. Magnetic source imaging supports clinical decision making in glioma patients. *Clinical neurology and neurosurgery* 2004;107:20-6.
185. Curra A, Modugno N, Inghilleri M, Manfredi M, Hallett M, Berardelli A. Transcranial magnetic stimulation techniques in clinical investigation. *Neurology* 2002;59:1851-9.

186. Krieg SM, Shibani E, Buchmann N, Meyer B, Ringel F. Presurgical navigated transcranial magnetic brain stimulation for recurrent gliomas in motor eloquent areas. *Clinical neurophysiology : official journal of the International Federation of Clinical Neurophysiology* 2013;124:522-7.
187. Picht T, Mularski S, Kuehn B, Vajkoczy P, Kombos T, Suess O. Navigated transcranial magnetic stimulation for preoperative functional diagnostics in brain tumor surgery. *Neurosurgery* 2009;65:93-8; discussion 8-9.
188. Negwer C, Sollmann N, Ille S, et al. Language pathway tracking: comparing nTMS-based DTI fiber tracking with a cubic ROIs-based protocol. *Journal of neurosurgery* 2017;126:1006-14.
189. Rosenstock T, Giampiccolo D, Schneider H, et al. Specific DTI seeding and diffusivity-analysis improve the quality and prognostic value of TMS-based deterministic DTI of the pyramidal tract. *NeuroImage Clinical* 2017;16:276-85.
190. Sollmann N, Wildschuetz N, Kelm A, et al. Associations between clinical outcome and navigated transcranial magnetic stimulation characteristics in patients with motor-eloquent brain lesions: a combined navigated transcranial magnetic stimulation-diffusion tensor imaging fiber tracking approach. *Journal of neurosurgery* 2018;128:800-10.
191. Frey D, Schilt S, Strack V, et al. Navigated transcranial magnetic stimulation improves the treatment outcome in patients with brain tumors in motor eloquent locations. *Neuro-oncology* 2014;16:1365-72.
192. Ille S, Sollmann N, Butenschoen VM, Meyer B, Ringel F, Krieg SM. Resection of highly language-eloquent brain lesions based purely on rTMS language mapping without awake surgery. *Acta neurochirurgica* 2016;158:2265-75.
193. Lu J, Wu J, Yao C, et al. Awake language mapping and 3-Tesla intraoperative MRI-guided volumetric resection for gliomas in language areas. *Journal of clinical neuroscience : official journal of the Neurosurgical Society of Australasia* 2013;20:1280-7.
194. Gasser T, Ganslandt O, Sandalcioglu E, Stolke D, Fahlbusch R, Nimsch C. Intraoperative functional MRI: implementation and preliminary experience. *NeuroImage* 2005;26:685-93.
195. Qiu TM, Gong FY, Gong X, et al. Real-Time Motor Cortex Mapping for the Safe Resection of Glioma: An Intraoperative Resting-State fMRI Study. *AJNR American journal of neuroradiology* 2017;38:2146-52.
196. Javadi SA, Nabavi A, Giordano M, Faghihzadeh E, Samii A. Evaluation of Diffusion Tensor Imaging-Based Tractography of the Corticospinal Tract: A Correlative Study With Intraoperative Magnetic Resonance Imaging and Direct Electrical Subcortical Stimulation. *Neurosurgery* 2017;80:287-99.
197. D'Andrea G, Familiari P, Di Lauro A, Angelini A, Sessa G. Safe Resection of Gliomas of the Dominant Angular Gyrus Availing of Preoperative FMRI and Intraoperative DTI: Preliminary Series and Surgical Technique. *World neurosurgery* 2016;87:627-39.
198. Dandy WE. Removal of right cerebral hemisphere for certain tumors with hemiplegia. *JAMA* 1928;90:823-5.
199. Duffau H. Long-term outcomes after supratotal resection of diffuse low-grade gliomas: a consecutive series with 11-year follow-up. *Acta neurochirurgica* 2016;158:51-8.

200. Yordanova YN, Moritz-Gasser S, Duffau H. Awake surgery for WHO Grade II gliomas within “noneloquent” areas in the left dominant hemisphere: toward a “supratotal” resection. Clinical article. *Journal of neurosurgery* 2011;115:232-9.
201. Verburg N, Pouwels PJ, Boellaard R, et al. Accurate Delineation of Glioma Infiltration by Advanced PET/MR Neuro-Imaging (FRONTIER Study): A Diagnostic Study Protocol. *Neurosurgery* 2016;79:535-40.
202. Dorward NL, Alberti O, Palmer JD, Kitchen ND, Thomas DG. Accuracy of true frameless stereotaxy: in vivo measurement and laboratory phantom studies. Technical note. *Journal of neurosurgery* 1999;90:160-8.
203. Mehta AD, Labar D, Dean A, et al. Frameless stereotactic placement of depth electrodes in epilepsy surgery. *Journal of neurosurgery* 2005;102:1040-5.
204. Holloway KL, Gaede SE, Starr PA, Rosenow JM, Ramakrishnan V, Henderson JM. Frameless stereotaxy using bone fiducial markers for deep brain stimulation. *Journal of neurosurgery* 2005;103:404-13.
205. Treuer H, Klein D, Maarouf M, Lehrke R, Voges J, Sturm V. Accuracy and conformity of stereotactically guided interstitial brain tumour therapy using I-125 seeds. *Radiotherapy and oncology : journal of the European Society for Therapeutic Radiology and Oncology* 2005;77:202-9.
206. Lonser RR, Warren KE, Butman JA, et al. Real-time image-guided direct convective perfusion of intrinsic brainstem lesions. Technical note. *Journal of neurosurgery* 2007;107:190-7.
207. Shamir RR, Joskowicz L, Spektor S, Shoshan Y. Target and trajectory clinical application accuracy in neuronavigation. *Neurosurgery* 2011;68:95-101; discussion -2.
208. Linskey ME. The changing role of stereotaxis in surgical neuro-oncology. *Journal of neuro-oncology* 2004;69:35-54.
209. Widmann G, Stoffner R, Sieb M, Bale R. Target registration and target positioning errors in computer-assisted neurosurgery: proposal for a standardized reporting of error assessment. *The international journal of medical robotics + computer assisted surgery : MRCAS* 2009;5:355-65.
210. Widmann G, Eisner W, Kovacs P, et al. Accuracy and clinical use of a novel aiming device for frameless stereotactic brain biopsy. *Minimally invasive neurosurgery : MIN* 2008;51:361-9.
211. Bernays RL, Kollias SS, Khan N, Romanowski B, Yonekawa Y. A new artifact-free device for frameless, magnetic resonance imaging-guided stereotactic procedures. *Neurosurgery* 2000;46:112-6; discussion 6-7.
212. Kelman C, Ramakrishnan V, Davies A, Holloway K. Analysis of stereotactic accuracy of the cosman-robert-wells frame and nexframe frameless systems in deep brain stimulation surgery. *Stereotactic and functional neurosurgery* 2010;88:288-95.
213. Fukaya C, Sumi K, Otaka T, et al. Nexframe frameless stereotaxy with multitract micro-recording: accuracy evaluated by frame-based stereotactic X-ray. *Stereotactic and functional neurosurgery* 2010;88:163-8.
214. Fitzpatrick JM, Konrad PE, Nickele C, Cetinkaya E, Kao C. Accuracy of customized miniature stereotactic platforms. *Stereotactic and functional neurosurgery* 2005;83:25-31.

215. Smith AP, Bakay RA. Frameless deep brain stimulation using intraoperative O-arm technology. Clinical article. *Journal of neurosurgery* 2011;115:301-9.
216. Varma TR, Eldridge PR, Forster A, et al. Use of the NeuroMate stereotactic robot in a frameless mode for movement disorder surgery. *Stereotactic and functional neurosurgery* 2003;80:132-5.
217. Mascott CR, Sol JC, Bousquet P, Lagarrigue J, Lazorthes Y, Lauwers-Cances V. Quantification of true in vivo (application) accuracy in cranial image-guided surgery: influence of mode of patient registration. *Neurosurgery* 2006;59:ONS146-56; discussion ONS-56.
218. Ortler M, Sohm F, Eisner W, et al. Frame-based vs frameless placement of intrahippocampal depth electrodes in patients with refractory epilepsy: a comparative in vivo (application) study. *Neurosurgery* 2011;68:881-7; discussion 7.
219. Cardinale F, Cossu M, Castana L, et al. Stereoelectroencephalography: surgical methodology, safety, and stereotactic application accuracy in 500 procedures. *Neurosurgery* 2013;72:353-66; discussion 66.
220. Dorfer C, Stefanits H, Pataraja E, et al. Frameless stereotactic drilling for placement of depth electrodes in refractory epilepsy: operative technique and initial experience. *Neurosurgery* 2014;10 Suppl 4:582-90; discussion 90-1.
221. Nowell M, Rodionov R, Diehl B, et al. A novel method for implementation of frameless StereoEEG in epilepsy surgery. *Neurosurgery* 2014;10 Suppl 4:525-33; discussion 33-4.
222. Ringel F, Ingerl D, Ott S, Meyer B. VarioGuide: a new frameless image-guided stereotactic system--accuracy study and clinical assessment. *Neurosurgery* 2009;64:365-71; discussion 71-3.
223. Hahn GJM, W.Q. *Statistical Intervals: A Guide for Practitioners*. New York: John Wiley & Sons; 1991.
224. Mascott CR. In vivo accuracy of image guidance performed using optical tracking and optimized registration. *Journal of neurosurgery* 2006;105:561-7.
225. Willems PW, Noordmans HJ, Ramos LM, et al. Clinical evaluation of stereotactic brain biopsies with an MKM-mounted instrument holder. *Acta neurochirurgica* 2003;145:889-97; discussion 97.
226. Westphal M, Lamszus K. The neurobiology of gliomas: from cell biology to the development of therapeutic approaches. *Nature reviews Neuroscience* 2011;12:495-508.
227. Omuro A, DeAngelis LM. Glioblastoma and other malignant gliomas: a clinical review. *JAMA* 2013;310:1842-50.
228. Harada K, Nishizaki T, Ozaki S, Kubota H, Ito H, Sasaki K. Intratumoral cytogenetic heterogeneity detected by comparative genomic hybridization and laser scanning cytometry in human gliomas. *Cancer Res* 1998;58:4694-700.
229. Little SE, Popov S, Jury A, et al. Receptor tyrosine kinase genes amplified in glioblastoma exhibit a mutual exclusivity in variable proportions reflective of individual tumor heterogeneity. *Cancer Res* 2012;72:1614-20.

230. van den Munckhof P, Contarino MF, Bour LJ, Speelman JD, de Bie RM, Schuurman PR. Postoperative curving and upward displacement of deep brain stimulation electrodes caused by brain shift. *Neurosurgery* 2010;67:49-53; discussion -4.
231. Chamberlain MC. Convection-enhanced delivery of a transforming growth factor-beta2 inhibitor trabedersen for recurrent high-grade gliomas: efficacy real or imagined?, in reference to Bogdahn et al. (*Neuro-Oncology* 2011;13:132-142). *Neuro-oncology* 2011;13:558-9; author reply 61-2.
232. Kubben PL, Wesseling P, Lammens M, et al. Correlation between contrast enhancement on intraoperative magnetic resonance imaging and histopathology in glioblastoma. *Surgical neurology international* 2012;3:158.
233. Stadlbauer A, Moser E, Gruber S, et al. Improved delineation of brain tumors: an automated method for segmentation based on pathologic changes of 1H-MRSI metabolites in gliomas. *NeuroImage* 2004;23:454-61.
234. Verburg N, Baayen JC, Idema S, et al. In vivo accuracy of a frameless stereotactic drilling technique for diagnostic biopsies and stereoelectroencephalography depth electrodes. *World neurosurgery* In press.
235. Sanai N, Berger MS. Glioma extent of resection and its impact on patient outcome. *Neurosurgery* 2008;62:753-64; discussion 264-6.
236. Stummer W, Reulen HJ, Meinel T, et al. Extent of resection and survival in glioblastoma multiforme: identification of and adjustment for bias. *Neurosurgery* 2008;62:564-76; discussion -76.
237. Grabowski MM, Recinos PF, Nowacki AS, et al. Residual tumor volume versus extent of resection: predictors of survival after surgery for glioblastoma. *Journal of neurosurgery* 2014;121:1115-23.
238. Marko NF, Weil RJ, Schroeder JL, Lang FF, Suki D, Sawaya RE. Extent of resection of glioblastoma revisited: personalized survival modeling facilitates more accurate survival prediction and supports a maximum-safe-resection approach to surgery. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2014;32:774-82.
239. Pan IW, Ferguson SD, Lam S. Patient and treatment factors associated with survival among adult glioblastoma patients: A USA population-based study from 2000-2010. *Journal of clinical neuroscience : official journal of the Neurosurgical Society of Australasia* 2015.
240. Favre J, Taha JM, Burchiel KJ. An analysis of the respective risks of hematoma formation in 361 consecutive morphological and functional stereotactic procedures. *Neurosurgery* 2002;50:48-56; discussion -7.
241. Kongkham PN, Knifed E, Tamber MS, Bernstein M. Complications in 622 cases of frame-based stereotactic biopsy, a decreasing procedure. *The Canadian journal of neurological sciences Le journal canadien des sciences neurologiques* 2008;35:79-84.
242. Institute NC. Common Terminology Criteria for Adverse Events v4.0 NIH publication # 09-7473 May 29, 2009.
243. Li J, Fine J. On sample size for sensitivity and specificity in prospective diagnostic accuracy studies. *Stat Med* 2004;23:2537-50.

244. Obuchowski NA. Sample size calculations in studies of test accuracy. *Statistical methods in medical research* 1998;7:371-92.
245. Obuchowski NA. Nonparametric analysis of clustered ROC curve data. *Biometrics* 1997;53:567-78.
246. Robin X, Turck N, Hainard A, et al. pROC: an open-source package for R and S+ to analyze and compare ROC curves. *BMC bioinformatics* 2011;12:77.
247. DeLong ER, DeLong DM, Clarke-Pearson DL. Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. *Biometrics* 1988;44:837-45.
248. Venkatraman ES. A permutation test to compare receiver operating characteristic curves. *Biometrics* 2000;56:1134-8.
249. Heiss P, Mayer S, Herz M, Wester HJ, Schwaiger M, Senekowitsch-Schmidtke R. Investigation of transport mechanism and uptake kinetics of O-(2-[<sup>18</sup>F]fluoroethyl)-L-tyrosine in vitro and in vivo. *Journal of Nuclear Medicine: Official Publication, Society of Nuclear Medicine* 1999;40:1367-73.
250. Lammertsma AA. Tracer Kinetic Modelling. In: Dierckx RAJO, Otte A, de Vries EFJ, van Waarde A, Leenders KL, eds. *PET and SPECT in Neurology*. Berlin, Heidelberg: Springer Berlin Heidelberg; 2014:59-73.
251. Langen K-J, Bartenstein P, Boecker H, et al. PET- und SPECT-Untersuchungen von Hirn tumoren mit radioaktiv markierten Aminosäuren. *Nuklearmedizin* 2011;50:167-73.
252. Vander Borght T, Asenbaum S, Bartenstein P, et al. EANM procedure guidelines for brain tumour imaging using labelled amino acid analogues. *European journal of nuclear medicine and molecular imaging* 2006;33:1374-80.
253. Bolcaen J, Lybaert K, Moerman L, et al. Kinetic Modeling and Graphical Analysis of 18F-Fluoromethylcholine (FCho), 18F-Fluoroethyltyrosine (FET) and 18F-Fluorodeoxyglucose (FDG) PET for the Discrimination between High-Grade Glioma and Radiation Necrosis in Rats. *PLOS ONE* 2016;11:e0161845.
254. Richard MA, Fouquet JP, Lebel R, Lepage M. Determination of an Optimal Pharmacokinetic Model of <sup>18</sup>F-FET for Quantitative Applications in Rat Brain Tumors. *Journal of Nuclear Medicine* 2017;58:1278-84.
255. Kratochwil C, Combs SE, Leotta K, et al. Intra-individual comparison of 18F-FET and 18F-DOPA in PET imaging of recurrent brain tumors. *Neuro-oncology* 2014;16:434-40.
256. Loeb R, Navab N, Ziegler SI. Direct Parametric Reconstruction Using Anatomical Regularization for Simultaneous PET/MRI Data. *IEEE Transactions on Medical Imaging* 2015;34:2233-47.
257. Thiele F, Ehmer J, Piroth MD, et al. The quantification of dynamic FET PET imaging and correlation with the clinical outcome in patients with glioblastoma. *Physics in Medicine and Biology* 2009;54:5525-39.
258. Pauleit D, Floeth F, Herzog H, et al. Whole-body distribution and dosimetry of O-(2-[<sup>18</sup>F]fluoroethyl)-l-tyrosine. *European journal of nuclear medicine and molecular imaging* 2003;30:519-24.



259. Zuhayra M, Alfteimi A, Forstner CV, Lutzen U, Meller B, Henze E. New approach for the synthesis of [18F]fluoroethyltyrosine for cancer imaging: simple, fast, and high yielding automated synthesis. *Bioorganic & medicinal chemistry* 2009;17:7441-8.
260. Gunn RN, Sargent PA, Bench CJ, et al. Tracer kinetic modeling of the 5-HT1A receptor ligand [carbonyl-11C]WAY-100635 for PET. *NeuroImage* 1998;8:426-40.
261. Boellaard R, Knaapen P, Rijbroek A, Luurtsema GJ, Lammertsma AA. Evaluation of basis function and linear least squares methods for generating parametric blood flow images using 15O-water and Positron Emission Tomography. *Mol Imaging Biol* 2005;7:273-85.
262. Gunn RN, Gunn SR, Cunningham VJ. Positron emission tomography compartmental models. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism* 2001;21:635-52.
263. Yaqub M, Boellaard R, Kropholler MA, Lammertsma AA. Optimization algorithms and weighting factors for analysis of dynamic PET studies. *Phys Med Biol* 2006;51:4217-32.
264. Akaike H. A new look at the statistical model identification. *IEEE Trans Autom Control* 1974;19(6):716-23.
265. Innis RB, Cunningham VJ, Delforge J, et al. Consensus nomenclature for in vivo imaging of reversibly binding radioligands. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism* 2007;27:1533-9.
266. Blomqvist G, Pauli S, Farde L, Eriksson L, Persson A, Halldin C. Maps of receptor binding parameters in the human brain--a kinetic analysis of PET measurements. *European journal of nuclear medicine* 1990;16:257-65.
267. Cunningham VJ, Hume SP, Price GR, Ahier RG, Cremer JE, Jones AK. Compartmental analysis of diprenorphine binding to opiate receptors in the rat in vivo and its comparison with equilibrium data in vitro. *Journal of cerebral blood flow and metabolism: official journal of the International Society of Cerebral Blood Flow and Metabolism* 1991;11:1-9.
268. Lammertsma AA, Hume SP. Simplified reference tissue model for PET receptor studies. *NeuroImage* 1996;4:153-8.
269. Bland JM, Altman DG. Measuring agreement in method comparison studies. *Statistical methods in medical research* 1999;8:135-60.
270. Langen KJ, Stoffels G, Filss C, et al. Imaging of amino acid transport in brain tumours: Positron emission tomography with O-(2-[(18F)fluoroethyl]-L-tyrosine (FET). *Methods* 2017;130:124-34.
271. Pauleit D, Floeth F, Herzog H, et al. Whole-body distribution and dosimetry of O-(2-[18F]fluoroethyl)-L-tyrosine. *European journal of nuclear medicine and molecular imaging* 2003;30:519-24.
272. Salinas CA, Searle GE, Gunn RN. The simplified reference tissue model: model assumption violations and their impact on binding potential. *Journal of cerebral blood flow and metabolism: official journal of the International Society of Cerebral Blood Flow and Metabolism* 2015;35:304-11.
273. Koopman T, Verburg N, Schuit RC, et al. Quantification of O-(2-[18F]fluoroethyl)-L-tyrosine kinetics in glioma. *EJNMMI Research* 2018;8.

274. Boellaard R, Knaapen P, Rijbroek A, Luurtsema GJJ, Lammertsma AA. Evaluation of Basis Function and Linear Least Squares Methods for Generating Parametric Blood Flow Images Using  $^{15}\text{O}$ -Water and Positron Emission Tomography. *Molecular Imaging and Biology* 2005;7:273-85.
275. Logan J, Fowler JS, Volkow ND, et al. Graphical Analysis of Reversible Radioligand Binding from Time – Activity Measurements Applied to [ $N$  -  $^{11}\text{C}$ -Methyl]-(-)-Cocaine PET Studies in Human Subjects. *Journal of Cerebral Blood Flow & Metabolism* 1990;10:740-7.
276. Cunningham VJ, Jones T. Spectral Analysis of Dynamic PET Studies. *Journal of Cerebral Blood Flow & Metabolism* 1993;13:15-23.
277. Logan J, Fowler JS, Volkow ND, Wang G-J, Ding Y-S, Alexoff DL. Distribution Volume Ratios without Blood Sampling from Graphical Analysis of PET Data. *Journal of Cerebral Blood Flow & Metabolism* 1996;16:834-40.
278. Gunn RN, Lammertsma AA, Hume SP, Cunningham VJ. Parametric Imaging of Ligand-Receptor Binding in PET Using a Simplified Reference Region Model. *NeuroImage* 1997;6:279-87.
279. Wu Y, Carson RE. Noise Reduction in the Simplified Reference Tissue Model for Neuroreceptor Functional Imaging. *Journal of Cerebral Blood Flow & Metabolism* 2002;22:1440-52.
280. Ichise M, Ballinger JR, Golan H, et al. Noninvasive quantification of dopamine D2 receptors with iodine-123-IBF SPECT. *Journal of Nuclear Medicine* 1996;37:513-20.
281. Ichise M, Liow J-S, Lu J-Q, et al. Linearized Reference Tissue Parametric Imaging Methods: Application to [ $^{11}\text{C}$ ]DASB Positron Emission Tomography Studies of the Serotonin Transporter in Human Brain. *Journal of Cerebral Blood Flow & Metabolism* 2003;23:1096-112.
282. Yaqub M, Tolboom N, Boellaard R, et al. Simplified parametric methods for [ $^{11}\text{C}$ ]PIB studies. *NeuroImage* 2008;42:76-86.
283. Bland JM, Altman DG. Measuring agreement in method comparison studies. *Statistical methods in medical research* 1999;8:135-60.
284. Ashburner J, Friston KJ. Unified segmentation. *NeuroImage* 2005;26:839-51.
285. Slifstein M, Laruelle M. Effects of statistical noise on graphic analysis of PET neuroreceptor studies. *Journal of Nuclear Medicine: Official Publication, Society of Nuclear Medicine* 2000;41:2083-8.
286. Ichise M, Cohen RM, Carson RE. Noninvasive Estimation of Normalized Distribution Volume: Application to the Muscarinic-2 Ligand [ $^{18}\text{F}$ ]FP-TZTP. *Journal of Cerebral Blood Flow & Metabolism* 2008;28:420-30.
287. Yaqub M, Tolboom N, van Berckel BNM, Scheltens P, Lammertsma AA, Boellaard R. Simplified parametric methods for [ $^{18}\text{F}$ ]FDDNP studies. *NeuroImage* 2010;49:433-41.
288. Jenkinson MD, Barone DG, Bryant A, et al. Intraoperative imaging technology to maximise extent of resection for glioma. *The Cochrane database of systematic reviews* 2018;1:CD012788.
289. Goldman S, Levivier M, Pirotte B, et al. Regional methionine and glucose uptake in high-grade gliomas: a comparative study on PET-guided stereotactic biopsy. *J Nucl Med* 1997;38:1459-62.

290. Pirotte B, Goldman S, Massager N, et al. Comparison of 18F-FDG and 11C-methionine for PET-guided stereotactic brain biopsy of gliomas. *J Nucl Med* 2004;45:1293-8.
291. Murphy RC, Kawashima A, Peller PJ. The utility of 11C-choline PET/CT for imaging prostate cancer: a pictorial guide. *AJR American journal of roentgenology* 2011;196:1390-8.
292. Chalaye J, Costentin CE, Luciani A, Nault JC. Reply to: "Response to: Positron emission tomography/computed tomography with (18)F-fluorocholine improve tumor staging and treatment allocation in patients with hepatocellular carcinoma". *Journal of hepatology* 2018;69:555-6.
293. Quak E, Blanchard D, Houdu B, et al. F18-choline PET/CT guided surgery in primary hyperparathyroidism when ultrasound and MIBI SPECT/CT are negative or inconclusive: the APACH1 study. *European journal of nuclear medicine and molecular imaging* 2018;45:658-66.
294. Hara T, Kondo T, Hara T, Kosaka N. Use of 18F-choline and 11C-choline as contrast agents in positron emission tomography imaging-guided stereotactic biopsy sampling of gliomas. *Journal of neurosurgery* 2003;99:474-9.
295. Kato T, Shinoda J, Nakayama N, et al. Metabolic assessment of gliomas using 11C-methionine, [18F] fluorodeoxyglucose, and 11C-choline positron-emission tomography. *AJNR American journal of neuroradiology* 2008;29:1176-82.
296. Mertens K, Ham H, Deblaere K, et al. Distribution patterns of 18F-labelled fluoromethylcholine in normal structures and tumors of the head: a PET/MRI evaluation. *Clinical nuclear medicine* 2012;37:e196-203.
297. Roelcke U, Bruehlmeier M, Hefti M, Hundsberger T, Nitzsche EU. F-18 choline PET does not detect increased metabolism in F-18 fluoroethyltyrosine-negative low-grade gliomas. *Clinical nuclear medicine* 2012;37:e1-3.
298. Spaeth N, Wyss MT, Pahnke J, et al. Uptake of 18F-fluorocholine, 18F-fluoro-ethyl-L-tyrosine and 18F-fluoro-2-deoxyglucose in F98 gliomas in the rat. *European journal of nuclear medicine and molecular imaging* 2006;33:673-82.
299. Kwee SA, Coel MN, Lim J, Ko JP. Combined use of F-18 fluorocholine positron emission tomography and magnetic resonance spectroscopy for brain tumor evaluation. *Journal of neuroimaging : official journal of the American Society of Neuroimaging* 2004;14:285-9.
300. Takenaka S, Asano Y, Shinoda J, et al. Comparison of (11)C-methionine, (11)C-choline, and (18)F-fluorodeoxyglucose-PET for distinguishing glioma recurrence from radiation necrosis. *Neurologia medico-chirurgica* 2014;54:280-9.
301. Spaeth N, Wyss MT, Weber B, et al. Uptake of 18F-fluorocholine, 18F-fluoroethyl-L-tyrosine, and 18F-FDG in acute cerebral radiation injury in the rat: implications for separation of radiation necrosis from tumor recurrence. *J Nucl Med* 2004;45:1931-8.
302. Verwer EE, Lammertsma AA, Boellaard R. Reply: Simplified Methods for Quantification of 18F-Fluoromethylcholine Uptake: Is SUVAUC,PP Actually an SUV? *J Nucl Med* 2015;56:1806-7.
303. Akaike H. Information theory and an extension of the maximum likelihood principle. *Selected Papers of Hirotugu Akaike*. Berlin: Springer; 1998:199-213.

304. Bossuyt PM, Reitsma JB, Bruns DE, et al. STARD 2015: an updated list of essential items for reporting diagnostic accuracy studies. *Radiology* 2015;277:826-32.
305. Bolcaen J, Descamps B, Deblaere K, et al. (18)F-fluoromethylcholine (FCho), (18)F-fluoroethyltyrosine (FET), and (18)F-fluorodeoxyglucose (FDG) for the discrimination between high-grade glioma and radiation necrosis in rats: a PET study. *Nuclear medicine and biology* 2015;42:38-45.
306. Lohmann P, Herzog H, Rota Kops E, et al. Dual-time-point O-(2-[(18)F]fluoroethyl)-L-tyrosine PET for grading of cerebral gliomas. *European radiology* 2015;25:3017-24.
307. Popperl G, Kreth FW, Mehrkens JH, et al. FET PET for the evaluation of untreated gliomas: correlation of FET uptake and uptake kinetics with tumour grading. *European journal of nuclear medicine and molecular imaging* 2007;34:1933-42.
308. Kracht LW, Friese M, Herholz K, et al. Methyl-[11C]-L-methionine uptake as measured by positron emission tomography correlates to microvessel density in patients with glioma. *European journal of nuclear medicine and molecular imaging* 2003;30:868-73.
309. Roivainen A, Forsback S, Gronroos T, et al. Blood metabolism of [methyl-11C]choline; implications for in vivo imaging with positron emission tomography. *European journal of nuclear medicine* 2000;27:25-32.
310. Ryken TC, Parney I, Buatti J, Kalkanis SN, Olson JJ. The role of radiotherapy in the management of patients with diffuse low grade glioma: A systematic review and evidence-based clinical practice guideline. *Journal of neuro-oncology* 2015;125:551-83.
311. Kinoshita M, Arita H, Goto T, et al. A novel PET index, 18F-FDG-11C-methionine uptake decoupling score, reflects glioma cell infiltration. *J Nucl Med* 2012;53:1701-8.
312. Gempt J, Soehngen E, Forster S, et al. Multimodal imaging in cerebral gliomas and its neuropathological correlation. *European journal of radiology* 2014;83:829-34.
313. Durst CR, Raghavan P, Shaffrey ME, et al. Multimodal MR imaging model to predict tumor infiltration in patients with gliomas. *Neuroradiology* 2014;56:107-15.
314. Hu LS, Ning S, Eschbacher JM, et al. Multi-Parametric MRI and Texture Analysis to Visualize Spatial Histologic Heterogeneity and Tumor Extent in Glioblastoma. *PLoS One* 2015;10:e0141506.
315. Chang PD, Malone HR, Bowden SG, et al. A Multiparametric Model for Mapping Cellularity in Glioblastoma Using Radiographically Localized Biopsies. *AJNR American journal of neuroradiology* 2017;38:890-8.
316. Fathi Kazerooni A, Nabil M, Zeinali Zadeh M, et al. Characterization of active and infiltrative tumorous subregions from normal tissue in brain gliomas using multiparametric MRI. *Journal of magnetic resonance imaging : JMRI* 2018.
317. Pallud J, Blonski M, Mandonnet E, et al. Velocity of tumor spontaneous expansion predicts long-term outcomes for diffuse low-grade gliomas. *Neuro-oncology* 2013;15:595-606.
318. Goze C, Blonski M, Le Maistre G, et al. Imaging growth and isocitrate dehydrogenase 1 mutation are independent predictors for diffuse low-grade gliomas. *Neuro-oncology* 2014;16:1100-9.

319. J. C. A coefficient of agreement for nominal scales. . Educational and Psychological Measurement 1960;20:37-46.
320. van Buuren S, Oudshoorn K. Flexible multivariate imputation by MICE. TNO Prevention and Health 1999.
321. Kracht LW, Miletic H, Busch S, et al. Delineation of brain tumor extent with [11C]L-methionine positron emission tomography: local comparison with stereotactic histopathology. Clinical cancer research: an official journal of the American Association for Cancer Research 2004;10:7163-70.
322. Filss CP, Galldiks N, Stoffels G, et al. Comparison of 18F-FET PET and perfusion-weighted MR imaging: a PET/MR imaging hybrid study in patients with brain tumors. J Nucl Med 2014;55:540-5.
323. Patel P, Baradaran H, Delgado D, et al. MR perfusion-weighted imaging in the evaluation of high-grade gliomas after treatment: a systematic review and meta-analysis. Neuro-oncology 2017;19:118-27.
324. Mandonnet E, Duffau H. An attempt to conceptualize the individual onco-functional balance: Why a standardized treatment is an illusion for diffuse low-grade glioma patients. Critical reviews in oncology/hematology 2018;122:83-91.
325. Wang M, Ma H, Wang X, et al. Integration of BOLD-fMRI and DTI into radiation treatment planning for high-grade gliomas located near the primary motor cortexes and corticospinal tracts. Radiation oncology 2015;10:64.
326. Eikenberry SE, Sankar T, Preul MC, Kostelich EJ, Thalhauser CJ, Kuang Y. Virtual glioblastoma: growth, migration and treatment in a three-dimensional mathematical model. Cell proliferation 2009;42:511-28.
327. Patel AP, Tirosh I, Trombetta JJ, et al. Single-cell RNA-seq highlights intratumoral heterogeneity in primary glioblastoma. Science 2014;344:1396-401.
328. Langen KJ, Hamacher K, Weckesser M, et al. O-(2-[18F]fluoroethyl)-L-tyrosine: uptake mechanisms and clinical applications. Nuclear medicine and biology 2006;33:287-94.
329. Koopman T, Verburg N, Schuit RC, et al. Quantification of O-(2-[(18)F]fluoroethyl)-L-tyrosine kinetics in glioma. EJNMMI Res 2018;8:72.
330. Faraway JJ. Extending the Linear Model with R: Generalized Linear, Mixed Effects and Nonparametric Regression Models, Second Edition, Edition 2: CRC Press
331. Bates D, Machler M, Bolker BM, Walker SC. Fitting Linear Mixed-Effects Models using lme4. Journal of Statistical Software 2015;67:1-48.
332. Buckley RC. Tissue Culture Studies of the Glioblastoma Multiforme. The American journal of pathology 1929;5:467-72 5.
333. Kim H, Zheng S, Amini SS, et al. Whole-genome and multisector exome sequencing of primary and post-treatment glioblastoma reveals patterns of tumor evolution. Genome research 2015;25:316-27.
334. Sottoriva A, Spiteri I, Piccirillo SG, et al. Intratumor heterogeneity in human glioblastoma reflects cancer evolutionary dynamics. Proceedings of the National Academy of Sciences of the United States of America 2013;110:4009-14.

335. Francis JM, Zhang CZ, Maire CL, et al. EGFR variant heterogeneity in glioblastoma resolved through single-nucleus sequencing. *Cancer discovery* 2014;4:956-71.
336. Nowell PC. The clonal evolution of tumor cell populations. *Science* 1976;194:23-8.
337. Mazor T, Pankov A, Song JS, Costello JF. Intratumoral Heterogeneity of the Epigenome. *Cancer cell* 2016;29:440-51.
338. Mazor T, Pankov A, Johnson BE, et al. DNA Methylation and Somatic Mutations Converge on the Cell Cycle and Define Similar Evolutionary Histories in Brain Tumors. *Cancer cell* 2015;28:307-17.
339. Wenger A, Ferreyra Vega S, Kling T, Bontell TO, Jakola AS, Caren H. Intratumor DNA methylation heterogeneity in glioblastoma: implications for DNA methylation-based classification. *Neuro-oncology* 2019;21:616-27.
340. Aryee MJ, Liu W, Engelmann JC, et al. DNA methylation alterations exhibit intraindividual stability and interindividual heterogeneity in prostate cancer metastases. *Science translational medicine* 2013;5:169ra10.
341. Ceccarelli M, Barthel FP, Malta TM, et al. Molecular Profiling Reveals Biologically Discrete Subsets and Pathways of Progression in Diffuse Glioma. *Cell* 2016;164:550-63.
342. Capper D, Jones DTW, Sill M, et al. DNA methylation-based classification of central nervous system tumours. *Nature* 2018;555:469-74.
343. Morrissy AS, Cavalli FMG, Remke M, et al. Spatial heterogeneity in medulloblastoma. *Nature genetics* 2017;49:780-8.
344. Wang Q, Hu B, Hu X, et al. Tumor Evolution of Glioma-Intrinsic Gene Expression Subtypes Associates with Immunological Changes in the Microenvironment. *Cancer cell* 2017;32:42-56 e6.
345. Benelli M, Romagnoli D, Demichelis F. Tumor purity quantification by clonal DNA methylation signatures. *Bioinformatics* 2018;34:1642-9.
346. Aran D, Sirota M, Butte AJ. Systematic pan-cancer analysis of tumour purity. *Nature communications* 2015;6:8971.
347. Zhang C, Cheng W, Ren X, et al. Tumor Purity as an Underlying Key Factor in Glioma. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2017;23:6279-91.
348. Prayson RA. The utility of MIB-1/Ki-67 immunostaining in the evaluation of central nervous system neoplasms. *Advances in anatomic pathology* 2005;12:144-8.
349. Reiter JG, Baretta M, Gerold JM, et al. An analysis of genetic heterogeneity in untreated cancers. *Nature reviews Cancer* 2019.
350. Gates EDH, Lin JS, Weinberg JS, et al. Guiding the first biopsy in glioma patients using estimated Ki-67 maps derived from MRI: conventional versus advanced imaging. *Neuro-oncology* 2019;21:527-36.
351. Nathanson V. Revising the Declaration of Helsinki. *Bmj* 2013;346:f2837.
352. McKnight TR, von dem Bussche MH, Vigneron DB, et al. Histopathological validation of a three-dimensional magnetic resonance spectroscopy index as a predictor of tumor presence. *Journal of neurosurgery* 2002;97:794-802.

353. Schiffer D, Valentini C, Melcarne A, et al. Spatial Relationships of MR Imaging and Positron Emission Tomography with Phenotype, Genotype and Tumor Stem Cell Generation in Glioblastoma Multiforme. In: Morgan LR, ed. Tumors of the Central Nervous System 2014.
354. Vander Borgh T, Asenbaum S, Bartenstein P, et al. EANM procedure guidelines for brain tumour imaging using labelled amino acid analogues. *European journal of nuclear medicine and molecular imaging* 2006;33:1374-80.
355. Van Cauter S, De Keyser F, Sima DM, et al. Integrating diffusion kurtosis imaging, dynamic susceptibility-weighted contrast-enhanced MRI, and short echo time chemical shift imaging for grading gliomas. *Neuro-oncology* 2014;16:1010-21.
356. Fudaba H, Shimomura T, Abe T, et al. Comparison of multiple parameters obtained on 3T pulsed arterial spin-labeling, diffusion tensor imaging, and MRS and the Ki-67 labeling index in evaluating glioma grading. *AJNR American journal of neuroradiology* 2014;35:2091-8.
357. Jena A, Taneja S, Jha A, et al. Multiparametric Evaluation in Differentiating Glioma Recurrence from Treatment-Induced Necrosis Using Simultaneous (18)F-FDG-PET/MRI: A Single-Institution Retrospective Study. *AJNR American journal of neuroradiology* 2017;38:899-907.
358. Zhang X, Rao A, Sette P, et al. IDH mutant gliomas escape natural killer cell immune surveillance by downregulation of NKG2D ligand expression. *Neuro-oncology* 2016;18:1402-12.
359. Siddiqui MM, Rais-Bahrami S, Turkbey B, et al. Comparison of MR/ultrasound fusion-guided biopsy with ultrasound-guided biopsy for the diagnosis of prostate cancer. *JAMA* 2015;313:390-7.
360. Friedewald SM, Rafferty EA, Rose SL, et al. Breast cancer screening using tomosynthesis in combination with digital mammography. *JAMA* 2014;311:2499-507.
361. Veit-Haibach P, Kuehle CA, Beyer T, et al. Diagnostic accuracy of colorectal cancer staging with whole-body PET/CT colonography. *JAMA* 2006;296:2590-600.
362. Bo HK, Solheim O, Jakola AS, Kvistad KA, Reinertsen I, Berntsen EM. Intra-rater variability in low-grade glioma segmentation. *Journal of neuro-oncology* 2017;131:393-402.
363. Unterrainer M, Vettermann F, Brendel M, et al. Towards standardization of (18)F-FET PET imaging: do we need a consistent method of background activity assessment? *EJNMMI Res* 2017;7:48.
364. Unkelbach J, Menze BH, Konukoglu E, Dittmann F, Ayache N, Shih HA. Radiotherapy planning for glioblastoma based on a tumor growth model: implications for spatial dose redistribution. *Phys Med Biol* 2014;59:771-89.
365. Rodriguez FJ, Giannini C. Chapter 7 - Diagnostic neuropathology of tumors of the central nervous system. In: Aminoff MJ, Boller F, Swaab DF, eds. *Handbook of Clinical Neurology* 2012.
366. Schneider M, Potthoff AL, Keil VC, et al. Surgery for temporal glioblastoma: lobectomy outranks oncosurgical-based gross-total resection. *Journal of neuro-oncology* 2019.
367. Burger PC, Heinz ER, Shibata T, Kleihues P. Topographic anatomy and CT correlations in the untreated glioblastoma multiforme. *Journal of neurosurgery* 1988;68:698-704.

368. Yamahara T, Numa Y, Oishi T, et al. Morphological and flow cytometric analysis of cell infiltration in glioblastoma: a comparison of autopsy brain and neuroimaging. *Brain tumor pathology* 2010;27:81-7.
369. van de Eede N, de Paepe A, Strens D, Specenier P. Real life cost of treatment and follow-up of patients with glioblastoma in Belgium: a retrospective patient chart review. *BELG J MED ONCOL* 2018;12:334-8.
370. De Witt Hamer PC, Ho VKY, Zwinderman AH, et al. Between-hospital variation in mortality and survival after glioblastoma surgery in the Dutch Quality Registry for Neuro Surgery. *Journal of neuro-oncology* 2019;144:313-23.



## II. Summary

This thesis set out to answer the following question: “Which MRI sequence, PET tracer, or combination of MRI sequence(s) and/or PET tracer(s) is the most accurate for the detection of diffuse glioma infiltration?”.

**Chapter 1** provides a general introduction, as well as the aim and outline of this thesis. After a description of the biological and clinical background of diffuse gliomas, we address the difficulty of the detection of diffuse glioma infiltration and the available imaging techniques for the detection of this infiltration. The aims of this thesis are: to discuss the current literature concerning imaging for the detection of diffuse glioma infiltration; to develop quantitative [ $^{18}\text{F}$ ]FET PET parametric maps; to compare the accuracy of different MRI sequences, PET tracers and combinations of MRI sequence(s) and/or PET tracer(s) for the detection of diffuse glioma infiltration; and to quantify such infiltration using histological, molecular and imaging techniques.

In **chapter 2** we systematically review the current literature on the diagnostic accuracy of imaging techniques for the detection of diffuse glioma infiltration. This meta-analysis on 61 studies, describing 3,532 samples in 1,309 patients, has several findings. First, the reporting quality of the studies is suboptimal. Second, the diagnostic accuracy for the detection of diffuse glioma infiltration of standard MRI used in daily practice is better for low-grade than high-grade glioma. Third, the diagnostic accuracy for the detection of diffuse glioma infiltration of standard MRI in high-grade gliomas is lower than MR spectroscopy and PET. Finally, samples without tumor presence from regions without imaging abnormalities (true negative samples) are underrepresented. Based on these findings, we conclude that a prospective study with direct comparison of imaging techniques including true negative samples is needed.

In **chapter 3** we discuss the literature on current available imaging techniques for the guidance of diffuse glioma resection and the localization of brain functions and white matter tracts. We conclude that, although multiple imaging techniques are available, studies directly comparing the guidance of diffuse glioma resection by different imaging techniques are sparse. The results of these few studies suggest that the use of other imaging techniques than the current standard MRI could result in improved survival for

patients. Another conclusion of this review is that direct cortical stimulation remains the gold standard for the localization of important brain functions and white-matter tracts. Imaging techniques, however, are indispensable for surgical planning, including the choice of awake versus non-awake surgery.

**Chapter 4** comprises a study of the accuracy of an in-house developed stereotactic drilling technique for diagnostic biopsies and stereo-electroencephalography depth electrode implantation. We included seven patients with 89 depth electrodes implanted and compared the pre-operative neuronavigation planning with the actual position on post-operative CT. The difference between planning and actual position was 3.5mm. Therefore, we conclude that our stereotactic drilling technique is suitable for diagnostic biopsies.

**Chapter 5** describes the study protocol for a monocenter prospective diagnostic accuracy study for the detection of diffuse glioma infiltration. Inclusion encompasses adult patients with a newly diagnosed, diffuse infiltrative glioma undergoing resective glioma surgery. Advanced neuroimaging is added to the standard preoperative MRI, and both are used to plan serial neuronavigated biopsies in and around the glioma boundaries. Biopsies are obtained immediately preceding resective surgery and provide histopathological and molecular characteristics of the regions of interest, enabling comparison with quantitative measurements in the imaging modalities at the same biopsy sites.

**Chapter 6** examines different tracer kinetic models for the quantification of [ $^{18}\text{F}$ ]FET kinetics in seven patients. The purpose of this study is to identify the optimal model using full plasma input data, as well as to identify the optimal simplified model that does not require arterial or venous blood sampling. The reversible two-tissue compartment model was the optimal tracer kinetic model and showed a strong correlation with the 60–90 min tumor-to-blood ratio, which was therefore identified as the optimal simplified model. There was also a significant, although moderate, correlation between [ $^{18}\text{F}$ ]FET kinetics and cerebral blood flow, determined by [ $^{15}\text{O}$ ]H<sub>2</sub>O PET. We conclude that the optimal simplified method is accurate enough to replace the full plasma input method.

**Chapter 7** compares multiple parametric maps of [ $^{18}\text{F}$ ]FET kinetics as well as tumor-to-brain ratio maps from different [ $^{18}\text{F}$ ]FET PET time intervals in seven patients. In order to assess the accuracy of the parametric maps, the mean value of the tumor volume was determined with the optimal model from chapter 6 and compared with the mean

value of the tumor volume of the different parametric maps. The quality of each image was assessed as the level of noise, with lower levels of noise representing higher image quality. The parametric maps based on the basis function method provided the best accuracy, while the parametric map based on the Logan method displayed the lowest level of noise. Tumor-to-normal ratio maps provided better accuracy and lower noise if later interval times were used.

**Chapter 8** consists of a study directly comparing [ $^{18}\text{F}$ ]FET and [ $^{11}\text{C}$ ]choline PET for the detection of diffuse glioma infiltration in 74 samples from eight patients. The diagnostic accuracy of both the standardized uptake value (SUV) and the tumor-to-brain ratio (TBR) maps of [ $^{11}\text{C}$ ]choline and different time intervals of [ $^{18}\text{F}$ ]FET were assessed. For [ $^{18}\text{F}$ ]FET, the diagnostic accuracy of the TBR was higher than that of the SUV for intervals 40–60 min and 60–90 min. For [ $^{11}\text{C}$ ]choline, there was no difference in diagnostic accuracy between the SUV and the TBR, however, there was a significant difference between tumor and normal samples SUV, but not TBR. The diagnostic accuracy of [ $^{18}\text{F}$ ]FET TBR 60–90 min was higher than that of [ $^{11}\text{C}$ ]choline SUV 20–40 min. We conclude that [ $^{18}\text{F}$ ]FET PET is more accurate than [ $^{11}\text{C}$ ]choline PET for detecting diffuse glioma infiltration.

**Chapter 9** presents the results of the direct comparison of diagnostic accuracy of multiple MRI sequences, PET tracers and combinations of MRI sequence(s) and/or PET tracer(s) for the detection of diffuse glioma infiltration in 174 samples from 20 patients. In enhancing gliomas, the combination of ADC with [ $^{18}\text{F}$ ]FET PET detected diffuse glioma infiltration better than T1G MRI and better than [ $^{18}\text{F}$ ]FET PET. In non-enhancing gliomas, no imaging combination detected diffuse glioma infiltration significantly better than standard MRI. FLAIR-weighted MRI was more accurate than [ $^{18}\text{F}$ ]FET PET in non-enhancing glioma. We constructed a probability map of tumor presence based on the combination of ADC with [ $^{18}\text{F}$ ]FET PET, with each voxel representing the probability of tumor presence, ranging from 0 to 100%. We conclude that combining ADC and [ $^{18}\text{F}$ ]FET PET detects diffuse glioma infiltration better than standard MRI and [ $^{18}\text{F}$ ]FET PET in enhancing gliomas, potentially enabling better local therapy.

**Chapter 10** quantifies diffuse glioma infiltration using different techniques and assesses intratumoral epigenetic heterogeneity in 133 samples of 16 patients. Diffuse glioma infiltration was quantified as tumor purity using (epi)genetic, histological and radiological metrics. An epigenetic tumor purity metric (PAMES) correlated best with all other metrics. Also, tumor purity demonstrated high inpatient spatial variation.

Intratumoral epigenetic heterogeneity was analyzed at molecular epigenetic classification level and compared with intratumoral variation in tumor purity. We conclude that apparent spatial heterogeneity in molecular epigenetic classification can generally be explained by variation in tumor purity and generally does not reflect biological variation. Genome-wide intratumoral epigenetic heterogeneity was also analyzed and confirmed the findings at molecular epigenetic classification level.

Finally, in **Chapter 11**, we present the general discussion of this thesis including: the current standard radiological imaging for the detection of diffuse glioma infiltration; imaging combinations for the detection of diffuse glioma infiltration; and quantification of diffuse glioma infiltration. Furthermore, we discuss the future perspectives of a new standard imaging protocol for diffuse glioma and local treatment beyond current standard MRI abnormalities. Finally, suggestions are made for future research.

## III. Nederlandse samenvatting

### (SUMMARY IN DUTCH)

Diffuse gliomen zijn de meest voorkomende hersentumoren. Ze ontstaan uit de (voorlopers van) gliacellen die als functie hebben de zenuwcellen (neuronen) in de hersenen te ondersteunen. De mate van kwaadaardigheid wordt uitgedrukt in een graderingssysteem van de Wereldgezondheidsorganisatie, waarbij graad I de minst en graad IV de meest kwaadaardige tumoren zijn. Diffuse gliomen zijn er in graad II, III en IV, waarbij graad IV glioblastoom wordt genoemd. Per jaar krijgen ongeveer 1100 mensen in Nederland een diffuus glioom, hiervan betreft het in circa 60% van de patiënten een glioblastoom.

De behandeling van diffuse gliomen bestaat uit een operatie en - afhankelijk van de graad, leeftijd en conditie van patiënt – bestraling en chemotherapie. Het doel van de operatie is om zo veel mogelijk tumorweefsel te verwijderen zonder ernstige neurologische uitval bij de patiënt te veroorzaken. De 10-jaars overleving is ongeveer 17%, hierdoor staan diffuse gliomen in de top vijf van meest dodelijke kankersoorten. Eén van de redenen dat diffuse gliomen zo moeilijk te behandelen zijn is hun neiging om het omliggende gezonde hersenweefsel wijdverspreid te infiltreren. Hierdoor is het lastig om tijdens een operatie onderscheid te maken tussen de tumor en normaal hersenweefsel. Om te helpen dit onderscheid te maken wordt er gebruik gemaakt van beeldvorming. De meest gebruikte techniek is beeldvorming met behulp van magnetische resonantie (MRI). MRI beelden kunnen op verschillende manieren verkregen worden, zogenaamde sequenties. De T2-, FLAIR- en T1 met contrast-gewogen sequenties zijn de huidige standaard voor het afbeelden van de infiltratie van een diffuus glioom. Er zijn ook andere MRI-sequenties en andere beeldvormende technieken, zoals bijvoorbeeld positronemissietomografie (PET) waarvoor meerdere radioactief gemerkte stoffen (tracers) bestaan, beschikbaar voor het afbeelden van deze tumorinfiltratie. Het is echter nog onvoldoende duidelijk of deze alternatieven beter zijn dan de huidige standaard.

Dit proefschrift heeft als doel het beantwoorden van de vraag: “Welke MRI sequentie, PET-tracer, of combinatie van MRI-sequentie(s) en/of PET-tracer(s), is het meest nauwkeurig voor het afbeelden van diffuusglioom-infiltratie?”.

**Hoofdstuk 1** betreft een algemene introductie en een korte beschrijving van het doel en de opzet van dit proefschrift. In de algemene discussie wordt eerst de biologische en klinische achtergrond van diffuse gliomen beschreven. Vervolgens komt de moeilijkheid van het detecteren van de infiltratie van diffuse gliomen aan de orde. De doelen van dit proefschrift zijn: bediscussiëren van de huidige literatuur omtrent beeldvorming voor het detecteren van de infiltratie van diffuus gliomen; ontwikkelen van PET beelden die gebaseerd zijn op metingen van de verdeling van de tracer [ $^{18}\text{F}$ ]FET, zogenaamde parametrische beelden; het vergelijken van de nauwkeurigheid van verschillende MRI-sequenties, PET-tracers, of combinatie van MRI-sequentie(s) en/of PET-tracer(s), voor het detecteren van de infiltratie van diffuse gliomen; en het kwantificeren van de diffuus glioom infiltratie met behulp van histologische, moleculaire en beeldvormende technieken.

**Hoofdstuk 2** bevat een systematische review van de literatuur over de diagnostische nauwkeurigheid van beeldvormende technieken voor het detecteren van diffuusglioom-infiltratie. Deze meta-analyse over 61 studies met 3,532 bipten in 1,309 patiënten leidde tot een aantal bevindingen. Ten eerste was de kwaliteit van rapporteren van de studies suboptimaal. Ten tweede, de diagnostische nauwkeurigheid voor het detecteren van diffuusglioom-infiltratie van standaard MRI, welke gebruikt wordt in de dagelijkse praktijk, is beter voor laaggradige dan hooggradige gliomen. Ten derde, de diagnostische nauwkeurigheid voor het detecteren van diffuusglioom-infiltratie van standaard MRI in hooggradige gliomen was minder dan die van magnetische resonantie spectroscopie en PET. Tot slot waren bipten zonder tumor uit gebieden zonder afwijkingen op de beeldvorming (echt negatieve bipten) ondervertegenwoordigd. Gebaseerd op deze bevindingen concluderen wij dat er een noodzaak is tot een prospectieve studie met directe vergelijking van beeldvormende technieken waarbij ook echt negatieve bipten worden verkregen.

In **hoofdstuk 3** bediscussiëren wij de literatuur omtrent de huidige beschikbare beeldvormende technieken voor het begeleiden van de resectie van diffuse gliomen en het lokaliseren van hersenfuncties en wittestofbanen. We concluderen dat er, ondanks de beschikbaarheid van meerdere beeldvormende technieken, weinig studies zijn die diffuusglioom-resecties op basis van verschillende beeldvormende technieken direct vergelijken. De resultaten van deze spaarzame studies suggereren dat het gebruik van andere beeldvormende technieken dan de huidige standaard MRI mogelijk zou kunnen leiden tot een verbeterde overleving van patiënten. Een andere conclusie is dat

directe corticale stimulatie de goudstandaard blijft voor het lokaliseren van belangrijke hersenfuncties en wittestofbanen. Beeldvormende technieken zijn echter onmisbaar voor de chirurgische planning, inclusief de keuze voor een wakkere, versus niet-wakkere, operatie.

**Hoofdstuk 4** bevat een studie naar de nauwkeurigheid van een stereotactische boortechniek, welke lokaal is ontwikkeld, voor diagnostische bipten en het inbrengen van stereo-electroencefalografische diepte-elektroden. We includeerden zeven patiënten met 89 ingebracht diepte-elektroden en vergeleken de preoperatieve neuro-navigatieplanning met de uiteindelijke positie op de CT. Het verschil tussen de planning en uiteindelijke positie was 3.5 mm. Daarom concluderen we dat onze stereotactische boortechniek geschikt is voor diagnostische bipten.

**Hoofdstuk 5** beschrijft het studieprotocol van een monocentrische prospectieve diagnostische nauwkeurigheid studie voor de detectie van diffuusgloom-infiltratie. Inclusie zal bestaan uit volwassen patiënten met een nieuwgediagnosticeerd diffuus gloom die een chirurgische resectie ondergaan. Geavanceerde beeldvormende technieken worden toegevoegd aan het standaard MRI-protocol. Beide worden gebruikt voor het plannen van seriële neurogenavigeerde bipten in, en rondom, de grenzen van het gloom. Bipten worden direct voorafgaande aan de resectie verkregen en gebruikt om de histopathologische en moleculaire kenmerken te bepalen. Dit maakt vergelijking met kwantitatieve metingen van de beeldvormende technieken van het biptie gebied mogelijk.

In **hoofdstuk 6** onderzoeken we verschillende kinetische modellen voor tracers om de  $[^{18}\text{F}]\text{FET}$ -kinetiek te kwantificeren in zeven patiënten. Het doel van deze studie was het identificeren van het optimale volledige model met behulp van de data van de bloedafnames, alsmede het identificeren van het optimale versimpelde model, welke bloedafnames overbodig maakt. Het reversibele twee-weefsel compartimentmodel was het optimale volledige tracerkinetische model en liet een sterke correlatie zien met de 60-90 min tumor-brein ratio, waardoor deze ratio als het optimale versimpelde model werd geïdentificeerd. Ook vonden we een matige maar significante correlatie tussen de  $[^{18}\text{F}]\text{FET}$ -kinetiek en cerebrale bloedstroom, zoals gemeten door middel van  $[^{15}\text{O}]\text{H}_2\text{O}$  PET. We concluderen dat de optimale versimpelde methode nauwkeurig genoeg is om de optimale volledige methode te vervangen.

In **hoofdstuk 7** vergelijken we in zeven patiënten meerdere [ $^{18}\text{F}$ ]FET PET parametrische beelden, alsmede tumor-brein ratio (TBR) beelden van verschillende [ $^{18}\text{F}$ ]FET PET intervallen. Om de nauwkeurigheid van de parametrische beelden te bepalen werd de gemiddelde waarde van het tumorvolume bepaald met behulp van de optimaal volledige methode uit hoofdstuk 6. Deze gemiddelde waarde werd vervolgens vergeleken met de gemiddelde waarde van het tumorvolume van de verschillende parametrische beelden. De kwaliteit van de beelden werd bepaald door de ruis te bepalen, waarbij minder ruis een betere beeldkwaliteit geeft. De parametrische beelden op basis van de basis functie methode waren het meest nauwkeurig. De parametrische beelden op basis van de Logan grafische analyse hadden de minste ruis. De nauwkeurigheid en ruis van de TBR-beelden waren beter voor de latere tijdsintervallen.

**Hoofdstuk 8** bevat een studie met een directe vergelijking van [ $^{18}\text{F}$ ]FET en [ $^{11}\text{C}$ ]choline PET voor de detectie van diffuusglio-infiltratie in 74 biopten van acht patiënten. De nauwkeurigheid van zowel de gestandaardiseerde opname waarde (SUV) als de TBR van [ $^{11}\text{C}$ ]choline en de verschillende tijdsintervallen van [ $^{18}\text{F}$ ]FET werden bepaald. Voor [ $^{18}\text{F}$ ]FET was de diagnostische nauwkeurigheid voor de detectie van diffuusglio-infiltratie van de TBR hoger dan van de SUV voor de tijdsintervallen 40–60min en 60–90min. Voor [ $^{11}\text{C}$ ]choline was er geen verschil in de diagnostische nauwkeurigheid voor de detectie van diffuusglio-infiltratie van de SUV en de TBR. Er was echter wel een significant verschil voor de SUV, maar niet voor de TBR, tussen tumor en normale samples. De diagnostische nauwkeurigheid van [ $^{18}\text{F}$ ]FET TBR 60–90min was hoger dan die van [ $^{11}\text{C}$ ]choline SUV 20–40min. We concluderen dat [ $^{18}\text{F}$ ]FET PET nauwkeuriger is dan [ $^{11}\text{C}$ ]choline PET voor de detectie van diffuusglio-infiltratie.

In **hoofdstuk 9** presenteren we de resultaten van de directe vergelijking van de diagnostische nauwkeurigheid van meerdere MRI-sequenties, PET-tracers, en combinaties van MRI-sequentie(s) en/of PET-tracer(s), voor het detecteren van diffuusglio-infiltratie in 174 biopten van 20 patiënten. In aankleurende gliomen bleek de combinatie van ADC met [ $^{18}\text{F}$ ]FET-PET de tumor beter te detecteren dan T1 met contrast-gewogen MRI en beter dan [ $^{18}\text{F}$ ]FET-PET. In niet-aankleurende gliomen was geen enkele combinatie beter in het detecteren van de tumor dan standaard MRI. FLAIR-gewogen MRI was nauwkeuriger dan [ $^{18}\text{F}$ ]FET-PET in niet-aankleurende gliomen. We construeerden een waarschijnlijkheidskaart voor de aanwezigheid van tumor, gebaseerd op de combinatie van ADC met [ $^{18}\text{F}$ ]FET-PET, waarbij elke voxel de waarschijnlijkheid van tumoraanwezigheid representeerde op een schaal van 0 tot 100%. We concluderen



dat in aankleurende gliomen de combinatie van ADC met [ $^{18}\text{F}$ ]FET-PET de mate van diffuusgliooin-infiltratie beter detecteert dan standaard MRI en [ $^{18}\text{F}$ ]FET-PET en dat dit kan leiden tot een betere lokale behandeling.

**Hoofdstuk 10** beschrijft een onderzoek waarin we diffuusgliooin-infiltratie met behulp van verschillende technieken kwantificeren en de intratumorale epigenetische heterogeniteit in 133 bipten van 16 patiënten bepalen. Diffuusgliooin-infiltratie werd gekwantificeerd als de mate van tumorpuurheid door middel van (epi)genetische, histologische en radiologische karakteristieken. Een epigenetische maat (PAMES) correleerde het beste met alle andere maten. Tumorpuurheid vertoonde een hoge mate van variatie binnen patiënten. Intratumorale epigenetische heterogeniteit werd geanalyseerd door de moleculaire epigenetische classificatie van alle samples te bepalen. Deze classificatie werd vervolgens vergeleken met de tumorpuurheid. We concludeerden dat ruimtelijke heterogeniteit in de moleculaire epigenetische classificatie meestal verklaard kan worden door de variatie in tumorpuurheid en niet het gevolg is van biologische variatie. Genoomwijde analyse van intratumorale epigenetische heterogeniteit bevestigde deze bevindingen.

In **hoofdstuk 11** presenteren we ten slotte de algemene discussie over de volgende onderwerpen: huidige standaard beeldvorming voor het detecteren van diffuusgliooin-infiltratie; combinaties van beeldvorming voor het detecteren van diffuusgliooin-infiltratie; en kwantificatie van diffuusgliooin-infiltratie. Tevens bediscussiëren we de toekomstperspectieven van een nieuw standaardprotocol voor beeldvorming van diffuse gliomen en lokale behandeling voorbij de afwijkingen op de huidige standaard-MRI. Op basis van deze discussie den we suggesties voor toekomstig onderzoek.

## IV. List of publications

### PUBLICATIONS INCLUDED IN THIS THESIS

Improved detection of diffuse glioma infiltration with imaging combinations: a diagnostic accuracy study.

**Verburg N**, Koopman T, Yaqub MM, Hoekstra OS, Lammertsma AA, Barkhof F, Pouwels PJW, Reijneveld JC, Heimans JJ, Rozemuller AJM, Bruynzeel AME, Lagerwaard F, Vandertop WP, Boellaard R, Wesseling P, de Witt Hamer PC.

Neuro Oncol. 2019 Sep 24. pii: noz180. doi: 10.1093/neuonc/noz180.

Direct comparison of [ $^{11}\text{C}$ ] choline and [ $^{18}\text{F}$ ] FET PET to detect glioma infiltration: a diagnostic accuracy study in eight patients.

**Verburg N**, Koopman T, Yaqub M, Hoekstra OS, Lammertsma AA, Schwarte LA, Barkhof F, Pouwels PJW, Heimans JJ, Reijneveld JC, Rozemuller AJM, Vandertop WP, Wesseling P, Boellaard R, de Witt Hamer PC.

EJNMMI Res. 2019 Jun 28;9(1):57. doi: 10.1186/s13550-019-0523-8.

Quantitative parametric maps of O-(2- $^{18}\text{F}$ fluoroethyl)-L-tyrosine kinetics in diffuse glioma.

Koopman T, **Verburg N**, Pouwels PJ, Wesseling P, Hoekstra OS, De Witt Hamer PC, Lammertsma AA, Yaqub M, Boellaard R.

J Cereb Blood Flow Metab. 2019 May 24:271678X19851878. doi: 10.1177/0271678X19851878.

Quantification of O-(2- $^{18}\text{F}$ fluoroethyl)-L-tyrosine kinetics in glioma.

Koopman T, **Verburg N**, Schuit RC, Pouwels PJW, Wesseling P, Windhorst AD, Hoekstra OS, de Witt Hamer PC, Lammertsma AA, Boellaard R, Yaqub M.

EJNMMI Res. 2018 Jul 31;8(1):72. doi: 10.1186/s13550-018-0418-0.

Diagnostic Accuracy of Neuroimaging to Delineate Diffuse Gliomas within the Brain: A Meta-Analysis.

**Verburg N**, Hoefnagels FWA, Barkhof F, Boellaard R, Goldman S, Guo J, Heimans JJ, Hoekstra OS, Jain R, Kinoshita M, Pouwels PJW, Price SJ, Reijneveld JC, Stadlbauer A, Vandertop WP, Wesseling P, Zwinderman AH, De Witt Hamer PC.

AJNR Am J Neuroradiol. 2017 Oct;38(10):1884-1891. doi: 10.3174/ajnr.A5368.

Accurate Delineation of Glioma Infiltration by Advanced PET/MR Neuro-Imaging (FRONTIER Study): A Diagnostic Study Protocol.

**Verburg N**, Pouwels PJ, Boellaard R, Barkhof F, Hoekstra OS, Reijneveld JC, Vandertop WP, Wesseling P, de Witt Hamer PC.

Neurosurgery. 2016 Oct;79(4):535-40. doi: 10.1227/NEU.0000000000001355.

In Vivo Accuracy of a Frameless Stereotactic Drilling Technique for Diagnostic Biopsies and Stereoelectroencephalography Depth Electrodes.

**Verburg N**, Baayen JC, Idema S, Klitsie MA, Claus S, de Jonge CS, Vandertop WP, de Witt Hamer PC.

World Neurosurg. 2016 Mar;87:392-8. doi: 10.1016/j.wneu.2015.11.041.

## OTHER PUBLICATIONS

Quantitative Third Harmonic Generation Microscopy for Assessment of Glioma in Human Brain Tissue.

Zhang Z, de Munck JC, **Verburg N**, Rozemuller AJ, Vreuls W, Cakmak P, van Huizen LMG, Idema S, Aronica E, de Witt Hamer PC, Wesseling P, Groot ML.

Adv Sci (Weinh). 2019 Apr 5;6(11):1900163. doi: 10.1002/advs.201900163. eCollection 2019 Jun 5.

Inter-rater agreement in glioma segmentations on longitudinal MRI.

Visser M, Müller DMJ, van Duijn RJM, Smits M, **Verburg N**, Hendriks EJ, Nabuurs RJA, Bot JCJ, Eijgelaar RS, Witte M, van Herk MB, Barkhof F, de Witt Hamer PC, de Munck JC.

Neuroimage Clin. 2019;22:101727. doi: 10.1016/j.nicl.2019.101727.

Effects of physician-based emergency medical service dispatch in severe traumatic brain injury on prehospital run time.

Franschman G, **Verburg N**, Brens-Heldens V, Andriessen TM, Van der Naalt J, Peerdeman SM, Valk JP, Hoogerwerf N, Greuters S, Schober P, Vos PE, Christiaans HM, Boer C.

Injury. 2012 Nov;43(11):1838-42. doi: 10.1016/j.injury.2012.05.020.

## V. Contributing authors

*In alphabetical order with affiliations and degrees at the time this research was conducted.*

**Kevin J. Anderson, Ph.D.**

Department of Computational Biology, Jackson Laboratory for Genomic Medicine, Farmington, USA

**Johannes C. Baayen, M.D.**

Department of Neurosurgery, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Frederik Barkhof, M.D. Ph.D.**

Department of Radiology & Nuclear Medicine, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

UCL institutes of Neurology & Healthcare Engineering, Gower St, Bloomsbury, London, United Kingdom

**Floris P. Barthel, M.D.**

Department of Computational Biology, Jackson Laboratory for Genomic Medicine, Farmington, USA

**Jeroen A.M. Belien, M.D. Ph.D.**

Department of Pathology, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Ronald Boellaard, Ph.D.**

Department of Radiology & Nuclear Medicine, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Anne M.E. Bruynzeel, M.D. Ph.D.**

Department of Radiotherapy, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Joseph F. Costello, Ph.D.**

Department of Neurological Surgery, UCSF, San Francisco, USA  
Karen Osney Brownstein Endowed Chair in Molecular Neuro-Oncology, UCSF,  
San Francisco, USA

**Steven Claus, M.D.**

Departement of Clinical Neurophysiology, Epilepsy Institutes in The Netherlands (SEIN),  
Heemstede, The Netherlands

**Serge Goldman, M.D. Ph.D.**

Service of Nuclear Medicine and PET/Biomedical Cyclotron Unit, l'universit'e libre de  
Bruxelles–Hôpital Erasme, Brussels, Belgium

**Jun Guo, M.D.**

Shanghai Medical College, Fudan University, Shanghai, China

**Jan J. Heimans, M.D. Ph.D.**

Department of Neurology, Amsterdam UMC, location VU medical center,  
Amsterdam, The Netherlands

**Friso W.A. Hoefnagels, M.D.**

Department of Neurosurgery, Amsterdam UMC, location VU medical center,  
Amsterdam, The Netherlands

**Otto S. Hoekstra, M.D. Ph.D.**

Department of Radiology & Nuclear Medicine, Amsterdam UMC, location VU medical  
center, Amsterdam, The Netherlands

**Sander Idema, M.D. Ph.D.**

Department of Neurosurgery, Amsterdam UMC, location VU medical center,  
Amsterdam, The Netherlands

**Rajan Jain, M.D. Ph.D.**

Department of Radiology, New York University School of Medicine, New York, New York

**Kevin C. Johnson, Ph.D.**

Department of Computational Biology, Jackson Laboratory for Genomic Medicine, Farmington, USA

**Catharina S. de Jonge, MSc**

Department of Neurosurgery, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Manabu Kinoshita, M.D. Ph.D.**

Department of Neurosurgery, Osaka University Graduate School of Medicine, Osaka, Japan

**Michiel A.J. Klitsie, MSc**

Department of Neurosurgery, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Frank Lagerwaard, M.D. Ph.D.**

Department of Radiotherapy, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Petra J.W. Pouwels, Ph.D.**

Department of Radiology & Nuclear Medicine, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Stephen J. Price, M.D. Ph.D.**

Academic Neurosurgery Division, Department of Clinical Neurosciences, Addenbrooke's Hospital, Cambridge, UK

**Annemieke J.M. Rozemuller, M.D. Ph.D.**

Department of Pathology, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Jaap C. Reijneveld, M.D. Ph.D.**

Department of Neurology, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Robert C. Schuit, BSc**

Department of Radiology & Nuclear Medicine, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Lothar A. Schwarte, M.D. Ph.D.**

Department of Anesthesiology, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands

**Andreas Stadlbauer, Ph.D.**

Department of Neurosurgery, University of Erlangen-Nuremberg, Erlangen, Germany

**Michael D. Taylor, M.D. Ph.D.**

Department of Neurosurgery, The Hospital for Sick Children, Toronto, Canada  
Departments of Surgery and Laboratory Medicine and Pathobiology,  
University of Toronto, Toronto, Canada

**W. Peter Vandertop, M.D. Ph.D.**

Department of Neurosurgery, Amsterdam UMC, location VU medical center,  
Amsterdam, The Netherlands  
Brain Tumor Center Amsterdam, Amsterdam UMC, Amsterdam, The Netherlands

**Roel G.W. Verhaak, Ph.D.**

Department of Computational Biology, Jackson Laboratory for Genomic Medicine,  
Farmington, USA

**Pieter Wesseling, M.D. Ph.D.**

Department of Pathology, Amsterdam UMC, location VU medical center,  
Amsterdam, The Netherlands  
Princess Máxima Centre for Paediatric Oncology, and Department of Pathology,  
University Medical Centre Utrecht, Utrecht, The Netherlands

**Albert D. Windhorst, Ph.D.**

Department of Radiology & Nuclear Medicine, Amsterdam UMC, location VU medical center, Amsterdam, The Netherlands



**Philip C. de Witt Hamer, M.D. Ph.D.**

Department of Neurosurgery, Amsterdam UMC, location VU medical center,  
Amsterdam, The Netherlands

Brain Tumor Center Amsterdam, Amsterdam UMC, Amsterdam, The Netherlands

**Aeilko H. Zwinderman, Ph.D.**

Department of Clinical Epidemiology and Biostatistics, Amsterdam UMC, location AMC,  
University of Amsterdam, Amsterdam, The Netherlands

## VI. Review Committee

**Prof. dr. B.M.J. Uitdehaag**

Department of Neurology, Amsterdam UMC, location VU medical center,  
Amsterdam, The Netherlands

**Prof. dr. R.G. Verhaak**

Department of Computational Biology, Jackson Laboratory for Genomic Medicine,  
Farmington, USA

**Prof. dr. G.A.M.S. van Dongen**

Department of Radiology and Nuclear Medicine, Amsterdam UMC, location VU medical  
center, Amsterdam, The Netherlands

**Prof. dr. M. Smits**

Brain Tumor Center, Rotterdam, The Netherlands  
Department of Radiology, Erasmus MC, Rotterdam, The Netherlands

**Dr. S.J. Price**

Academic Neurosurgery Division, Department of Clinical Neurosciences,  
Addenbrooke's Hospital, Cambridge, UK

**Dr. A.M.E. Walenkamp**

Department of Medical Oncology, University Medical Center Groningen,  
Groningen, The Netherlands

## VII. PhD Portofolio

Name student: **N. Verburg**  
 PhD period: January 2016 – January 2020  
 PhD supervisors: prof. dr. W.P. Vandertop  
 prof. dr. P. Wesseling  
 prof. dr. P.C. de Witt Hamer

	Organizer	Year	EC
<b>Mandatory</b>			
Scientific integrity course	VUmc Academy	2018	2.0
Participation and presentation at international conferences	Society of Neuro Oncology	2016-2019	4.0
Participation and presentation at national conferences	Dutch Neuro Oncology workgroup meeting, Dutch Radiology Meeting, Dutch Neurosurgery Wintermeeting	2016-2019	4.0
Statistics course	Exemption		2.0
Oncology course	Exemption		3.0
<b>Subtotal</b>			<b>15.0</b>
<b>Elective Mandatory</b>			
Participation in weekly clinical oncology meetings	VUmc	2016-2019	2.0
Participation in weekly journal clubs	VUmc	2016-2019	4.0
Organization of journal clubs	VUmc	2016-2019	2.0
Tutor honors program student	VU	2016	2.0
Tutor student	VU	2019	2.0
Writing scientific articles under supervision	VUmc	2016-2019	2.0
Education for (scrub) nurses	Amstel Academy	2016-2018	2.0
Writing grant application	Niels Stensen Fellowship <i>Awarded</i>	2019	2.0
<b>Subtotal</b>			<b>18.0</b>
<b>Elective</b>			
Radiation hygiene	Boerhaave	2019	1.7
R2 introduction workshop	AMC Onco Genomics	2016	0.5
<b>Subtotal</b>			<b>2.2</b>
<b>Total</b>			<b>35.2</b>

## VIII. List of dissertations Brain Tumor Center Amsterdam

Date	PhD Candidate	Thesis
10-06-2005	Carla Verstappen	Cancer therapy related neurotoxicity
28-09-2005	Maaïke Vos	Evaluation of response, toxicity and outcome in glioma therapy
20-12-2005	Birgit Georger	Conditionally replicative adenoviruses for the treatment of malignant glioma and neuroblastoma
20-12-2005	Jacques Grill	Gene therapy and virotherapy of brain tumors with recombinant adenoviruses
19-06-2009	Fonnet Bleeker	Mutational profiling of glioblastoma
24-11-2009	Philip de Witt Hamer	Glioblastoma: between bed and bench
07-05-2010	Ingeborg Bosma	Cognitive dysfunction in glioma; underlying mechanisms and consequences
23-09-2010	Christian Badr	Bioluminescence imaging in glioblastoma: monitoring of biological processes and novel therapeutics
08-11-2010	Linda Douw	Neural networks in brain tumors; the interplay between tumor, cognition, and epilepsy
10-06-2011	Sander Idema	Improving oncolytic viral therapy for glioma
05-10-2011	Anneke Niers	Novel biosensors for preclinical brain tumor analysis
16-11-2011	Krista van Niftrik	In vitro studies on radiation and temozolomide in human glioma
03-07-2012	Viola Caretti	Pioneering preclinical research in diffuse intrinsic pontine glioma: towards new treatment strategies
29-10-2012	Leonora Balaj	Exosomes: the biological messengers
08-02-2013	Marjolein de Groot	Epilepsy in brain tumor patients; towards improved and personalized treatment
04-06-2013	Edwin van Dellen	Lesions in the connected brain; a network perspective on brain tumors and lesional epilepsy
04-12-2013	Michiel Smits	Micro-RNA and epigenetic signaling in glioma angiogenesis
11-12-2013	Eefje Sizoo	The end-of-life phase of high-grade glioma patients; towards a dignified death
17-06-2014	Dannis van Vuurden	Innovative treatment targets in pediatric high-grade brain tumors

Date	PhD Candidate	Thesis
07-01-2015	Lotte Hiddingh	Treatment sensitizers for high-grade tumors
17-03-2015	Marc Jansen	Diffuse Intrinsic Pontine Glioma: clinical aspects and imaging
11-05-2015	Florien Boele	Towards improving health-related quality of life in glioma patients and their informal caregivers
30-09-2015	Johan Koekkoek	Epilepsy in glioma patients; optimizing treatment until the end of life
19-01-2016	Sjoerd van Rijn	Functional molecular imaging of cancer development and stem cell regeneration in the nervous system
24-03-2016	Hinke van Thuijl	Molecular characterization of low-grade glial neoplasms
18-05-2016	Avanita Prabowo	Molecular Features of long-term epilepsy-associated tumours; focus on glioneural tumours
08-06-2016	Ronald Willemse	Functional mapping of the sensorimotor cortex: clinical studies with MEG and fMRI
21-06-2016	Sharyar Mir	Novel treatment targets in high-grade brain tumors
01-07-2016	Femke Froklage	The role of the blood-brain barrier in drug-resistance and central neurotoxicity
03-11-2016	Thijs Crommentuijn	Development of vector-based strategies against glioblastoma
28-11-2017	Lot Sewing	Diffuse Intrinsic Pontine Glioma: disease models and translational research
28-11-2017	Sophie Veldhuijzen van Zanten	Diffuse Intrinsic Pontine Glioma: a multi-facetted and global view
22-12-2017	Remco Molenaar	Radiation in cancer. From cellular to clinical effects
23-01-2018	Tonny Lagerweij	Brain tumors: Preclinical imaging and novel therapies
14-09-2018	Ravi Narayan	Radiosensitizing and synergistic targeted therapy for glioblastoma
27-09-2018	Friso Hoefnagels	Imaging techniques as an adjunct in resective brain surgery
23-01-2020	Niels Verburg	Advanced imaging in glioma treatment; moving the frontier.

## IX. Dankwoord

Ibiza, zomer 2018: “Sorry lief, maar ik moet nu even doorwerken want dan is het dit jaar nog allemaal klaar. Dat wil jij toch ook?” Dus... ruim een jaar en twee vakanties verder, waar helaas ook in gewerkt werd, is het nu dan écht zover. Wat begon met een idee over een onderzoekje naar fluorescerende middelen als coassistent heeft uiteindelijk geleid tot dit proefschrift aan het einde van mijn opleiding tot neurochirurg. Zoals iedereen weet is dit natuurlijk geen solo project en hebben er velen geholpen die ik graag wil bedanken.

Allereerst natuurlijk alle **patiënten**, en hun **families**, die kostbare tijd en energie belangeloos hebben gegeven voor dit project. Het is bijna niet voor te stellen in wat voor situatie deze mensen zitten. Net geconfronteerd met een ernstige ziekte en met het vooruitzicht van een hersenoperatie, hebben zij toch uren extra onder de scanner gelegen en zijn bereid geweest om een langere procedure te ondergaan. Mijn grootste hoop is dat we met het onderzoek écht hebben bijgedragen aan het verbeteren van de behandeling van mensen met een diffuus glioom.

Mijn promotor, **prof. dr. W.P. Vandertop**. Beste professor, voor een promovendus met haast is bijna niet te beschrijven hoe prettig het was dat elk stuk binnen no-time, voorzien van zeer scherp en accuraat commentaar, terug kwam in mijn inbox. De precisie met betrekking tot de Nederlandse en Engelse taal heeft mij niet alleen geholpen in het schrijven van artikelen en beursaanvragen, maar ook nog eens met het behalen van het IELTS Engels examen. Dezelfde nauwkeurigheid is er op de OK en in het klinisch redeneren, waar ik enorm veel van heb geleerd. Daarnaast heb ik mij altijd gesteund gevoeld in het combineren van opleiding en promotie. Bedankt voor de flexibiliteit in het inzetten van mijn KWF beurs. Hopelijk kunnen we in de toekomst nog veel samenwerken. De (twee?)jaarlijkse assistenten BBQ's ga ik missen!

Mijn promotor, **prof. dr. P. Wesseling**. Beste Pieter, zonder jou was dit boekje er nooit gekomen. Jouw kennis, inzicht en kritische blik hebben mij enorm geholpen. Ondanks je vele functies en neventaken, maakte je altijd tijd voor mij vrij. Mede dankzij jouw uitgebreide hulp lukte het om de KWF beurs te verkrijgen. Daarnaast kwam ik door jou in contact met Roel Verhaak en recent heb je weer enorm bijgedragen aan het verkrijgen van het Niels Stensen Fellowship. Mailtjes rond middernacht, telefoontjes

op zondagochtend, altijd sta je voor me klaar. Naast mooie academische kwaliteiten, ben je ook oprecht geïnteresseerd in de mensen om je heen en weet je iedereen te waarderen (en dit ook te benoemen). Ik hoop in de toekomst nog veel met je te kunnen samenwerken en biertjes op vliegvelden te drinken na SNO meetings.

Mijn copromotor, **prof. dr. P.C. de Witt Hamer**. Beste Philip, jij hebt mij wegwijs gemaakt in de wetenschap. En zoals je dat met alles doet, ging dit gepaard met een ongelooflijk analytisch vermogen en enorm perfectionisme. Dit heeft mij erg geholpen en ik heb ontzettend veel van je geleerd. Het is knap hoe jij naast je klinische werk een onderzoeksgroep heb weten op te richten, wat zeer terecht heeft geleid tot een hoogleraarschap. Een cursus time management heb jij niet meer nodig, 15 minuten introductie uitleg in R moet genoeg zijn toch? Naast onderzoek doen, heb ik ook veel van je geleerd op de OK en de polikliniek. Waarschijnlijk besef je wel dat we nog veel werk te verzetten hebben, aangezien er nog een whisky & sigaren avond gepland moet worden en er nog een halve doos FRONTIER wijn op jouw kamer ligt. O ja, misschien moeten we ook nog éven die RCT opstarten waar we het al jaren over hebben...

**Prof. dr. B.M.J. Uitdehaag, prof. dr. G.A.M.S. van Dongen, prof. dr. M. Smits en dr. A.M.E. Walenkamp**. Hartelijk dank voor de tijd en moeite die u heeft genomen om mijn proefschrift te beoordelen en uw bereidheid om zitting te nemen in de promotiecommissie. Excuses voor het stalken door zowel promotor als promovendus om snel het oordeel te vellen, wat zelfs doorging tot op Curaçao. Zonder u was het nooit gelukt om nog voor mijn start in Cambridge mijn proefschrift te verdedigen.

**Prof. dr. R. G. Verhaak**. Beste Roel, mijn periode in jouw lab heb ik als zeer inspirerend en leerzaam ervaren. Een nieuwe wereld ging voor mij open en jij hebt me de kans gegeven deze zelf te ontdekken. De manier waarop jij je onderzoeksgroep leidt en weet te motiveren is voor mij een voorbeeld. Je hebt langer dan je gewend bent moeten wachten op de resultaten van onze samenwerking. Het nadeel van een onderzoeker die ook nog in opleiding is tot neurochirurg. Nu die opleiding voorbij is durf je het misschien weer aan om in de toekomst nieuwe gezamenlijke projecten aan te gaan. En ik beloof om je de volgende keer niet te laten zitten bij de 'presidential reception' op de SNO meeting.

**Dr. S. J. Price**. There are many things to thank you for. It starts with you sharing your data with us, which led to the meta-analysis. Next, I would like to thank you for your time and effort of reviewing my thesis in such a short time. Finally, I am very thankful for the great

opportunity you have given me by allowing me to join your group in Cambridge for my Niels Stensen Fellowship. I am looking forward to our collaboration.

De FRONTIER onderzoeksgroep, **prof. dr. F. Barkhof, prof. dr. O.S. Hoekstra, prof. dr. R. Boellaard, prof. dr. J.J. Heimans, dr. P.J.W. Pouwels, dr. M.M. Yaqub, dr. J. Reijneveld** en **T. Koopmans**. Beste Frederik, Otto, Ronald, Jan, Petra, Maqsood, Jaap en Thomas, zonder jullie was de FRONTIER studie nooit geslaagd. Het was een uitdaging om met al onze verschillende achtergronden dezelfde taal te spreken. Toch is dit gelukt en heeft het tot mooie resultaten geleid. Jullie hebben veel bijgedragen aan mijn wetenschappelijke vorming waarvoor dank. En nu op naar het vervolg van de FRONTIER!

The Verhaak crew, **Kevin, Floris, K2, Samir** and **Hoon**. You made me feel really welcome at the Jackson Lab. Besides extending my R and basic data analyses skills, you have thought me a lot about the American culture: drinking IPA, watching superbowl, not to drive in a snowblizzard and courtyard beerpong. I am looking forward to future SNO meetings.

**Prof. dr. A.J.M. Rozemuller**. Beste Annemieke, bedankt voor al je tijd en moeite die je hebt gestoken in het beoordelen, en soms nogmaals gezamenlijk beoordelen, van alle samples.

**Dr. J.A.M. Belien**. Beste Jeroen, zonder jou was ik nu nog steeds bezig geweest met het bepalen van de MIB index. Eén van mijn grootste onderschattingen van dit project. Met behulp van jouw pipeline kon de enorme hoeveelheid data toch verwerkt worden. Dank hiervoor.

De CCA laboranten, **Dirk, François** en **Paul**. Zonder jullie enorme inspanning was het nooit gelukt om het moleculaire project op de rails te krijgen. Onder grote tijdsdruk hebben jullie topwerk geleverd. Daarnaast hebben we geleerd dat Molfix toch niet de beste keuze was...

Alle **PET en MRI laboranten**. Dit logistiek méér dan uitdagende project was nooit gelukt zonder jullie inzet. Ondanks problemen met de tracers, de meerdere scans op één dag en ook nog eens ontelbaar veel bloedsamples hebben we het voor elkaar gekregen. Het creatief plannen van **Judith** was onmisbaar. Bedankt allemaal.

Beste **Gem**, bedankt voor het afnemen van bloedmonsters bij patiënten.



Alle **OK-assistenten** van de neurochirurgie. 174 biopten neem je niet zonder een goed team. Jullie hulp bij dit uitdagende onderdeel was geweldig!

PICTURE groep, **Domenique, Roeland** en **Martin**. Het is nooit helemaal gelukt om de FRONTIER en PICTURE bij elkaar te krijgen, maar aan jullie heeft het niet gelegen. Nog elke week moet ik de PICTURE meeting afspraak in mijn agenda weigeren. Vaak zijn jullie een klankbord geweest waarvoor dank. Hopelijk komen binnenkort de eerste PICTURE boekjes uit.

Mijn opleider, **prof. dr. S. M. Peerdeman**. Beste Saskia, zonder jouw flexibiliteit in mijn opleidingsschema was het allemaal nooit gelukt. Daarnaast heb je mij altijd goed geholpen om mijn focus te houden ondanks de vele andere interessante projecten die op mijn pad kwamen. Met dit boekje komt ook een einde aan mijn opleiding. Ik heb enorm veel van je geleerd, zowel op OK als daarbuiten. De manier waarop jij kliniek en management weet te combineren is een voorbeeld. Ik ga de jaarlijkse AIOS etentjes bij jou thuis erg missen.

Stafleden Amsterdam UMC, locatie Vumc. Beste **Hans, David, Ronald, Ricardo** en **Sander**. Dank voor jullie onuitputtelijke kritische academische blik die altijd op zeer subtiële wijze kenbaar werd gemaakt. Ondanks het feit dat jullie je de helft van de tijd je afvroegen waar ik was en wat ik in vredesnaam aan het doen was, heb ik toch veel van jullie geleerd. Voornamelijk slechte grappen, koffie drinken en wat skiën. Maar gelukkig ook nog opereren. Ik heb het enorm naar mijn zin gehad op de Boelelaan, Mahler en de verschillende Alpenretraites. Ik hoop nog veel met jullie samen te kunnen werken, bedankt!

Stafleden Amsterdam UMC, locatie AMC. Beste **Gert Joan, Rick, Bert, Michiel, Pepijn, Dennis, Friso, Jantien, Mariam, Maarten** en **René**. Ik heb mijn altijd zeer welkom gevoeld in het AMC en het OLVG West. Ook jullie hebben je vaak afgevraagd waar ik nu weer was en waarom ik nog steeds niet klaar was met de opleiding. Ik heb veel van jullie geleerd op de onderdelen die we op de locatie Boelelaan niet zo vaak zien. Ook met jullie hoop ik nog veel samen te kunnen werken in de toekomst, bedankt!

Mede-AIOS **Rob, Paul, Ivar, George, Roos, Hisse, Steven, Anil** en **Yasmine**. Eindelijk is het zover, niet meer die eeuwige AIOS als chieft resident. Ik besef dat mijn exorbitant lange opleiding niet altijd even fijn voor jullie is geweest, maar gelukkig was ik er vaak ook

niet. Jullie begrijpen natuurlijk wel dat ik me enigszins zorgen maak over de aankomende Alpenretraite, aangezien ik jullie niet meer kan chaperonneren. Ditzelfde geldt natuurlijk voor de NVNA cursussen, waarbij ik wel verwacht dat de Amsterdamse academische eer hoog gehouden wordt. Ik kijk ernaar uit om in de toekomst met jullie samen te werken.

ANIOS locatie VUmc. Beste **Myron, Yvette, Victoria** en **Sophie**. Bedankt dat jullie de echte Heroes of the Ward zijn!

Secretariaat neurochirurgie Amsterdam UMC, locatie VUmc. Beste **Wendy, Ingrid** en **Brigitte**. Bedankt voor al het zichtbare en onzichtbare werk.

OK technicus **Walter**. Bedankt voor al je hulp bij de verschillende technische uitdagingen van deze studie en alle andere projecten.

Alle **verpleegkundigen, medisch secretaressen** en **voedingsassistenten** van het neurocentrum Amsterdam UMC, locatie VUmc. Bedankt voor de goede zorg voor de patiënten en altijd fijne sfeer op de afdeling.

De wijnsnobs **Jurjen, Peter, Willem, Reinier, Jeroen, Rob** en **Rogier**. Ondanks dat we van schrale pils over zijn gegaan op pretentieuze Bordeauxs en we in plaats van vier dagen per week elkaar vier uur per maand zien, is er verder weinig veranderd. Geen grap wordt onbenut gelaten. Bedankt voor de broodnodige afleiding en ik hoop elkaar weer vaker te kunnen zien.

Bonifanten **Rutger, Arthur, Iris** en **Loekie**. Wat bijzonder om nog steeds bevriend te zijn sinds de eerste klas middelbare school. Ik hecht hier veel waarde aan en hoop ook jullie in de toekomst weer vaker te kunnen zien. Helaas voor jullie is Cambridge net zo duur en ongezeellig als Amsterdam...

Mijn paranimf **Bart**. Gezien onze symbiose kon dit boekje niet uitblijven. Vanaf nu mag ik weer aan dezelfde tafel zitten als onze vrouwen en jou. Naast onze geweldige en voor mij zeer belangrijke vriendschap, ben jij ook wetenschappelijk een voorbeeld met je prachtige onderzoekscarrière tot nu toe. Misschien dat we in de toekomst nog een keer samen kunnen publiceren. En als het allemaal niet lukt hebben we altijd nog onze HNP bus om op terug te vallen.

Mijn paranimf **Jacob**. Ons werk als (neuro)chirurg, gezamenlijke fanatisme in spelletjes en voorliefde voor feestjes zijn een rode draad in onze vriendschap. De vele vakanties in het huis van jouw oom in Perletto hebben de wijnsnobs gevormd. Hoewel we nooit samen hebben geopereerd, was het OK complex wel de plek om even bij te praten in onze blauwe pakken. Geen biertjes meer bij Mahler voor ons, maar wie weet waar wel in de toekomst.

Lieve **Ien** en **Willem**. Bedankt voor jullie constante oprechte interesse. Jullie zijn als familie voor me.

Lieve **Flip**. Bedankt voor jouw kritische en analytische denken, gecombineerd met een perfectionisme in taal, waarmee je mij enorm hebt geholpen bij meerdere brieven en het essay voor de aanvraag van mijn Niels Stensen Fellowship.

Lieve **Menso, Tourette** en **Maria**. Ik ben ontzettend blij met jullie als schoonfamilie. Altijd zeer betrokken en erg geïnteresseerd in wat ik doe. Daarnaast een enorme hulp als oppas van Feline en Pepijn wat regelmatig nodig was met al mijn nationale en internationale verplichtingen. In de toekomst gelukkig weer meer tijd om Sinterklaas voor te bereiden.

Lieve oma, lieve **Rie**. Helaas mocht jij niet studeren van je vader. Ik weet zeker dat je een voortreffelijk wetenschapper had kunnen worden met jouw onuitputtelijke nieuwsgierigheid. Jammer dat Lex niet meer bij mijn promotie kan zijn. Echter was dit misschien ook niet helemaal zijn ding, aangezien hij me na het horen van mijn KWF beurs vroeg of ik niet eerst een keer moest gaan werken voordat ik weer ging studeren. Bedankt dat er iets van jouw nieuwsgierigheid aan mij is doorgegeven.

Lieve pap en mam, lieve **Dick** en **Sjan**. Tijdens het schrijven van het essay voor het Niels Stensen Fellowship besepte ik mij weer goed dat jullie mij gemaakt hebben tot wie ik ben en dat ik hier enorm dankbaar voor ben. Jullie idealisme is bewonderingswaardig en een voorbeeld voor me. Ik heb mij altijd enorm gesteund gevoeld in de keuzes in mijn leven, waaronder ook dit proefschrift. Daarnaast hebben jullie ook enorm geholpen door op te passen op Feline en Pepijn. Nu er weer meer tijd is hoop ik jullie vaker te zien en te spreken.

Lieve **Feline** en **Pepijn**, misschien vinden jullie ooit een ondergestofte doos vol met boekjes op zolder en bladeren jullie er doorheen. Weet dat ik alles eraan heb gedaan om dit proefschrift niet in de weg te laten staan van mijn tijd met jullie. Dus gooi het boekje maar snel terug in de doos en dan gaan we iets leuks doen.

Mijn lief, **Hannah**. Van iedereen heb jij het meeste moeten verduren van mijn promotie. De vele avonden dat ik met een laptop op de bank zat, jij hoogzwanger van Pepijn alleen voor Feline moeten zorgen toen ik in het lab in Amerika was en mijn afwezigheid tijdens de internationale congressen. Ondanks dit alles ga je ook nog met mij mee naar Cambridge. Er zijn geen woorden hiervoor, maar ik dank je voor alles wat je hebt gedaan. Weet dat jij en de kinderen het allerbelangrijkste in mijn leven zijn. Ik hou van je!

En nu is het tijd voor een klein drankje...!

## X. Curriculum Vitae

Niels Verburg was born on September 11, 1984 in Utrecht, The Netherlands. After obtaining his atheneum diploma at the St. Bonifatiuscollege in Utrecht in 2002, he worked as a volunteer in a primary school and hospital in Ghana, Africa. In 2003 he started to study Medicine at the Free University, Amsterdam. Between his study and internships he travelled to South and Central America. An internship in Emergency Medicine was followed at the Kalafong Hospital in Pretoria, South Africa. After graduating as a medical doctor in 2010, he started as a resident not in training at the department of Neurosurgery, VU Medical Center, Amsterdam. In 2012 his neurosurgical training commenced at the department of Neurosurgery, VU medical center, Amsterdam (chair: prof. dr. W.P. Vandertop) under guidance of prof. dr. S.M. Peerdeman. He trained for 6 months in spinal surgery under guidance of dr. G.J. Bouma at the OLVG West, Amsterdam. His training has completed in December 2019.

In 2011 he started his collaboration with prof. dr. P.C. de Witt Hamer, who just started a research group for diffuse glioma imaging. After receiving a grant of the Cancer Center Amsterdam in 2012, they initiated the FRONTIER study. In 2015, Niels obtained a Dutch Cancer Foundation Resident Grant, which allowed him to combine his residency with PhD research under guidance of prof. dr. P.C. de Witt Hamer, prof. dr. W.P. Vandertop and prof. dr. P. Wesseling. In 2017, he spent two months as a visiting scientist in the Computational Biology laboratory of prof. dr. R.J.W. Verhaak at the Jackson Laboratory for Genomic Medicine (Farmington, USA), which has led to an ongoing collaboration. The results of this collaboration and his PhD research are presented in this thesis. Recently he was awarded a Niels Stensen Fellowship that allows him to continue his research at the University of Cambridge, UK under guidance of dr. S.J. Price. Niels will be starting this new chapter in his life together with his wife Hannah and their two children Feline and Pepijn.



